

ALGAL OIL: A NOVEL SOURCE OF OMEGA-3 FATTY ACIDS FOR HUMAN NUTRITION

Osman Kadir TOPUZ

Seafood Processing Technology Department, Fisheries Faculty of Akdeniz University, Pinarbasi
Mah. Konyaalti, Antalya, Turkey, Phone: +90242 310 60 19, Email: oktopuz@akdeniz.edu.tr

Corresponding author email: oktopuz@akdeniz.edu.tr

Abstract

Docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) a polyunsaturated fatty acid (PUFA) that belongs to the omega-3 fatty acids group. In recent years, omega-3 fatty acids rich oil has attracted much attention because of its recognized beneficial effect on human health. At present, fish oil is the major source of omega-3 fatty acids, but omega-3 fatty acids rich oils can be produced by microalgae with additional commercial benefits. Consumption of omega-3 rich fish oil has been steadily increasing for decades due to their health benefits. Microalgal oil might be the most promising alternative to fish oil, since they are the primary producers of omega-3 fatty acids. Recent advances regarding production of omega-3 fatty acid rich oil from microalgae have been reviewed with an emphasis on the scientific data. At first, description of the omega-3 fatty acids, biosynthesis pathways and their role in the human health is presented. Microalgae are the initial source of omega-3 fatty acids. Microalgae species intensively used in omega-3 fatty acids rich algal oil production and their culturing conditions were reviewed in this paper. The algal oil extraction and refining process are also presented. We present here a review of the most recent advances regarding the production of omega-3 fatty acid rich algal oil from the marine origin microalgae.

Key words: Omega-3 fatty acids, docosahexaenoic acid, eicosapentaenoic acid, microalgae, algal oil.

INTRODUCTION

Polyunsaturated fatty acids (PUFA), specifically docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) have been linked to several health benefits (Armenta & Valentine, 2013; Covington, 2004). Several studies have suggested there is positive health effects consumption of DHA and EPA, such as lowering the risk of vision loss due to eye macular degeneration, reduction of blood vessel stiffness, and relief of anxiety and inflammation (Ho, van Leeuwen, Witteman, van Duijn, Uitterlinden, Hofman, et al., 2011; Kiecolt-Glaser, Belury, Andridge, Malarkey, & Glaser, 2011; Pase, Grima, & Sarris, 2011). DHA consumption has been associated with reducing risk of colds in babies from moms that took this dietary supplement (Imhoff-Kunsch, Stein, Martorell, Parra-Cabrera, Romieu, & Ramakrishnan, 2011). All the latter, plus numerous studies that suggest there are health effects for the heart and brain, and many other benefits for treating several illnesses and diseases including asthma, rheumatoid arthritis, schizophrenia, depression, multiple sclerosis,

migraine headaches, etc (Gray, 2011). Even low DHA levels have been associated to suicide risk (Armenta & Valentine, 2013; Lewis, Hibbeln, Johnson, Lin, Hyun, & Loewke, 2011).

The major sources of EPA and DHA in food and dietary supplements were found in fatty fish, fish products, marine oils, and certain algae oils (Andız & Ünlüsayın 2015). At present, the major resource of omega-3 fatty acids for human nutrition is marine fish oils. However, their reserves are dwindling, and supplies via wild fish fisheries are limited. Moreover, omega-3 fatty acid content of oil from wild-caught fish varies with species, location, water depth and temperature, seasonal climatic conditions, and availability and type of primary food chain (Khozin-Goldberg, Iskandarov, & Cohen, 2011). A variety of fish species such as herring, mackerel, sardine and salmon are regarded as good sources of omega-3 fatty acids. Due to the many shortcomings of fish-derived oil including undesirable taste and odour, diminishing supplies, objections by vegetarians, its chemical processing methods, and the presence of contaminants such as

mercury, dioxins and polychlorinated biphenyls (Certik & Shimizu, 1999; Hooper, Thompson, Harrison, Summerbell, Ness, Moore, et al., 2006), research has been diverted towards the exploitation of other marine species for the development of a suitable and sustainable alternatives (Gupta, Barrow, & Puri, 2012). Microalgae offer a promising non-polluted resource for biotechnology and bioengineering of omega-3 fatty acid oil production, as an alternative to fish oil. Compared to terrestrial crop plants, microalgae present a few advantages as omega-3 fatty acids sources, such as commonly occurring genes for the biosynthesis of these nutrients, simpler acid profiles and higher growth rates (D. A. Martins, Custodio, Barreira, Pereira, Ben-Hamadou, Varela, et al., 2013). Most of oil producing microalgae studied within the past decade have been eukaryotes; and have been found worldwide, both along coastlines and in the open ocean. Using microalgae to produce omega-3 fatty acid rich algal oil is still a relatively new field, and research on this area has been growing significantly within the last few years (Armenta & Valentine, 2013). Studied microalgae include *Nannochloropsis oculata* (Pal, Khozin-Goldberg, Cohen, & Boussiba, 2011) *Pavlova lutheri*, *Odontella aurita* (Guihéneuf, Fouqueray, Mimouni, Ulmann, Jacquette, & Tremblin, 2010), *Schizochytrium* sp. (I. Fedorova-Dahms, P. A. Marone, M. Bauter, & A. S. Ryan, 2011), *Cryptocodinium cohnii* (Mendes, Reis, Vasconcelos, Guerra, & da Silva, 2009), *Ulkenia* sp. (Quilodrán, Hinzpeter, Hormazabal, Quiroz, & Shene, 2010). This review focus on recent advances made in biotechnological production of omega-3 fatty acids rich oil from microalgae alternative to fish oil.

Omega-3 fatty acids

Polyunsaturated fatty acids (PUFAs) constitute a large group of fatty acids containing long chain carbonic molecules that include ω -3 and ω -6 fatty acids. Omega ' ω ' is the position of the first double bond when counted from the methyl end and the number '3' refers to the number of carbon atoms at that position from the methyl end. The molecular structure of the fatty acids consists of an even number of carbon atoms (4 to 24) with diverse saturations

(0 to 6 double bonds) (Gupta, Barrow, & Puri, 2012). Docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA) are two members of the ω -3 family (Figure 1).

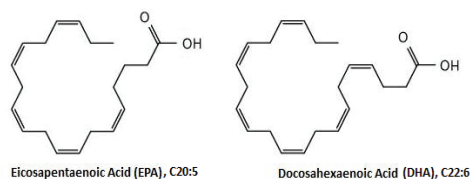


Figure 1. Structure of common omega-3 fatty acids: DHA and EPA.

Biosynthesis pathways of EPA and DHA are shown in Figure 2. The prefixes 'docosa' and 'eicosa' are of Greek descent, meaning the 22 and 20 C atoms present in DHA and EPA, which contain 6 and 5 cis-double bonds respectively (Lozac'h, 1986). PUFAs rich in omega-6 fatty acids originate from terrestrial plants are consumed in higher abundance than those from fish, so that plant origin ω -6 fatty acids are consumed in higher quantities than ω -3 fatty acids (Newton, 1998). This over consumption of ω -6 relative to ω -3 oil has been linked to increased risk of cancer, diabetes, cardiovascular and neurodegenerative diseases (A. Simopoulos, 2006). To restore a balance, consumption of ω -3 fatty acids should be increased compared to ω -6 fatty acids. ω -6 to ω -3 PUFA ratios of between 5:1 and 3:1 have been suggested as optimum for human consumption (A. P. Simopoulos, 2008).

Omega-3 fatty acids rich microalgae

In the marine food system, polyunsaturated fatty acids are primarily formed by phytoplankton and transferred on to herbivorous zooplankton, hence affecting food quality for organism at higher trophic levels. Various photoautotrophic and heterotrophic marine species from different classes produce EPA and DHA omega-3 fatty acids.

According to recent reviews of total lipid extracts, *Bacillariophyceae* (diatoms) and *Chrysophyceae* species may be rich sources of EPA and DHA; *Cryptophyceae*, *Prasinophyceae*, *Rhodophyceae*, *Xanthophyceae*, *Glaucophyceae* and *Eustigmatophyceae* can represent interesting

EPA sources, whereas DHA is found in significant amounts mostly in *Dinophyceae*, *Prymnesiophyceae*, and *Euglenophyceae* (Lang, Hodac, Friedl, & Feussner, 2011; D. A. Martins, et al., 2013).

These organisms offer a promising vegetative and non-polluted resource for biotechnology and bioengineering of omega-3 fatty acids rich algal oil production as an alternative to fish oil.

Currently, the production of omega-3 fatty acids by marine microorganisms is the subject of intensive research and increasing commercial attention (L. Sijtsma & M. E. de Swaaf, 2004). Several photosynthetic and heterotrophic marine microalgae are considered as a good source of omega-3 fatty acids (EPA and DHA) for the commercial production of algal oil (Table 1).

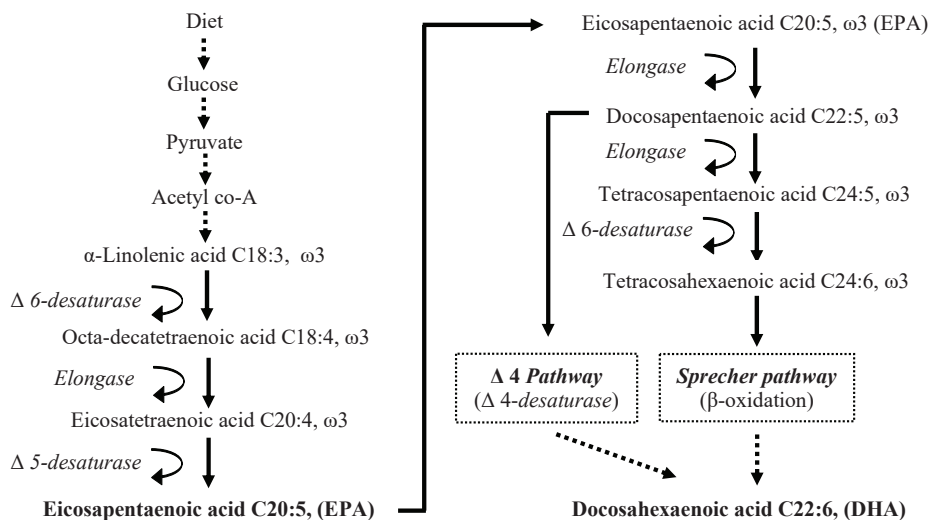


Figure 2. Biosynthesis pathways of EPA and DHA ω -3 fatty acids.

Photoautotrophic microalgae

It is generally thought that photosynthetic microalgae tend to produce higher levels of EPA than heterotrophs. *Nannochloropsis* sp. Hibberd, *Phaeodactylum* Bohlin, *Nitzschia* Hassall and *Porphyridium* Nägeli can present elevated levels of EPA in total fatty acids, although relatively low cell lipid contents tend to result in small EPA amounts in the biomass (D. A. Martins, et al., 2013) (Table 1). *Pavlova lutheri* and *Nannochloropsis oculata* are marine microalgae commonly used in aquaculture, is a well-known source of n-3 LC-PUFA, such as EPA and DHA under specific conditions such as during exponential growth using low light (Guihéneuf, Mimouni, Tremblin, & Ulmann, 2015; Winwood, 2013).

Heterotrophic microalgae

Currently, the most common micro-algae used for the production of DHA rich algal oil are

from the marine members of the families *Thraustochytriaceae* and *Cryptecodiniaceae*.

The Thraustochytrids include the genera *Schizochytrium* sp. and *Ulkenia* sp., whereas *Cryptecodinium* is a genus of the family Cryptecodiniaceae (Table 1). Thraustochytrids, which includes the genera *Schizochytrium* and *Thraustochytrium*, are among the most promising microorganisms for producing omega-3 fatty acids, with reported oil contents of >50 % (dry basis) and more than 30 % DHA within the total oil produced (Burja, Radianingtyas, Windust, & Barrow, 2006). Among the heterotrophic marine microalgae, *Cryptecodinium cohnii* was identified as a prolific producer of DHA. The *C. cohnii* is extraordinary in that it produces no other PUFAs other than DHA in its cell lipid in any significant amount (Mendes, Reis, Vasconcelos, Guerra, & da Silva, 2009; Van Pelt, Huang, Tschanz, & Brenna, 1999).

Table 1. Microalgae species intensively used in omega-3 fatty acids rich algal oil production

Microalgae	Culture condition	Omega-3 fatty acids	References
<i>Schizochytrium</i> sp.	Heterotrophic	24-45 % DHA, 10 % EPA	(I. Fedorova-Dahms, P. Marone, M. Bauter, & A. Ryan, 2011; Hammond, Mayhew, Naylor, Ruecker, Mast, & Sander, 2001)
<i>Ulkenia</i> sp.	Heterotrophic	35-40 % DHA	(Dulce Alves Martins, Custódio, Barreira, Pereira, Ben-Hamadou, Varela, et al., 2013)
<i>Crypthecodinium cohnii</i>	Heterotrophic	25-60 % DHA	(De Swaaf, Sijtsma, & Pronk, 2003; L. Sijtsma & M. De Swaaf, 2004)
<i>Thraustochytrium striatum</i>	Heterotrophic	37 % DHA, 23 % EPA	(Fan, Chen, Jones, & Vrijmoed, 2001)
<i>Aurantiochytrium</i> sp.	Heterotrophic	40 % DHA	(Hong, Rairakhwada, Seo, Park, Hur, Kim, et al., 2011)
<i>Pavlova lutheri</i>	Phototrophic	12-18 % DHA, 22-28 % EPA	(Guihéneuf, Mimouni, Ulmann, & Tremblin, 2009)
<i>Nannochloropsis</i> sp.	Phototrophic	38-39% EPA	(Chaturvedi & Fujita, 2006)
<i>Phaeodactylum tricornutum</i>	Phototrophic	40-57 % EPA	(Fernández, Pérez, Sevilla, Camacho, & Grima, 2000)
<i>Nitzschia laevis</i>	Phototrophic	25-33 % EPA	(Xiao-Hong, Song-Yao, WANG, & Mei-Fang, 2007)
<i>Porphyridium cruentum</i>	Phototrophic	25 % EPA	(Durmaz, Monteiro, Bandarra, Gökpinar, & Işik, 2007)

Extraction and purification of algal oil

After growth and harvesting of microbial biomass, oil must be gently extracted. Nonetheless, there is no universal method that will yield the best oil recovery for algae. In some algae cells, cell walls are particularly thick and cell disruption method must be employed for recovery process. Extraction of algal oil for the production of omega 3 fatty acids is difficult process because as soon as the algal cell walls are ruptured, these PUFAs are exposed to potential oxidation. Once these highly unsaturated fatty acids have reacted with oxidized radicals, an unstoppable chain reaction begins which leads to the production of rancid, highly odorous oil which is unsuitable for human consumption. Hence, so far as possible, all sources of materials that can initiate the oxidation process should be eliminated during the extraction and storage period (Winwood, 2013). Crude vegetable oils, including algal oils, require refining to improve color, clarity, odour and remove any particulate material and chemical contaminants. There are wide ranges of impurities in the crude oil that can be removed by the refining process, including: free fatty acids, phosphatides (i.e. lecithin), pigments (i.e. carotenoids, chlorophyll), trace metals, sterols (i.e. cholesterol), waxes, mono acyl and diacyl

glycerides, waxes, oxidation products and trace contaminants. Therefore, oil refining needs to be performed to remove non-triglyceride, colorants, smelly and toxic compounds in the production of edible oil (Rubio-Rodríguez et al., 2010). The conventional oil refining in industry is usually made by chemical methods, which include several steps as degumming, to separate phospholipids; neutralization or de-acidification, to clear free fatty acids and decrease oil acidity; bleaching to absorb pigments or contaminants and deodorization to remove smelly compounds. Supercritical fluid technology, together with membrane and enzymatic processes, is one of the most recent technologies proposed as alternative to oil refining with chemical products or high temperatures (Čmolík & Pokorný, 2000).

CONCLUSIONS

Scientists have a great opportunity to discover and exploit many as yet unidentified marine microbes capable of producing higher levels of omega-3 fatty acids and other valuable products. One of the main challenges is developing optimum culture conditions for rapidly growing marine microbes that produce high levels of omega-3 fatty acids, as compared to the limited number of commercially useful

species presently available in the international depository. The application of metabolic engineering techniques to improve the wild-type strain is in an early stage of development. The bioengineering can further improve yields of omega-3 fatty acids from algae. Consumers are aware of the importance of an adequate provision of these nutrients and several properties of microalgal oils are particularly appealing, such as their sustainability, high purity and quality, “vegetarian” origin, and improved organoleptic qualities when compared to fish oils. Although genetically modified crops will likely serve as omega-3 sources in the future, microalgae oils have a great potential to present purer profiles, which are highly advantageous during processing and may address differentiated purposes in the market.

ACKNOWLEDGEMENTS

The authors would like to express their sincere thanks to Scientific Research Project Coordination Unit of Akdeniz University and the Scientific and Technological Research Council of Turkey (TUBİTAK project code: 214O721) for the support.

REFERENCES

- Armenta, R. E., Valentine, M. C. 2013. Single-Cell Oils as a Source of Omega-3 Fatty Acids: An Overview of Recent Advances. *Journal of the American Oil Chemists Society*, 90 (2), 167-182.
- Burja, A. M., Radianingtyas, H., Windust, A., Barrow, C. J. 2006. Isolation and characterization of polyunsaturated fatty acid producing *Thraustochytrium* species: screening of strains and optimization of omega-3 production. *Applied Microbiology and Biotechnology*, 72 (6), 1161-1169.
- Certik, M., Shimizu, S. 1999. Biosynthesis and regulation of microbial polyunsaturated fatty acid production. *Journal of Bioscience and Bioengineering*, 87 (1), 1-14.
- Chaturvedi, R., Fujita, Y. 2006. Isolation of enhanced eicosapentaenoic acid producing mutants of *Nannochloropsis oculata* ST-6 using ethyl methane sulfonate induced mutagenesis techniques and their characterization at mRNA transcript level. *Phycological Research*, 54 (3), 208-219.
- Covington, M. B. 2004. Omega-3 fatty acids. *American Family Physician*, 70 (1), 133-140.
- De Swaaf, M. E., Sijtsma, L., Pronk, J. T. 2003. High-cell-density fed-batch cultivation of the docosahexaenoic acid producing marine alga *Cryptocodinium cohnii*. *Biotechnology and Bioengineering*, 81 (6), 666-672.
- Durmaz, Y., Monteiro, M., Bandarra, N., Gökpinar, Ş., Işık, O. 2007. The effect of low temperature on fatty acid composition and tocopherols of the red microalga, *Porphyridium cruentum*. *Journal of Applied Phycology*, 19(3), 223-227.
- Fan, K., Chen, F., Jones, E. B., Vrijmoed, L. L. 2001. Eicosapentaenoic and docosahexaenoic acids production by and okara-utilizing potential of *thraustochytrids*. *Journal of Industrial Microbiology and Biotechnology*, 27(4), 199-202.
- Fedorova-Dahms, I., Marone, P., Bauter, M., Ryan, A. 2011. Safety evaluation of DHA-rich Algal Oil from *Schizochytrium* sp. *Food and Chemical Toxicology*, 49 (12), 3310-3318.
- Fernández, F., Pérez, J., Sevilla, J., Camacho, F. G., Grima, E. M. 2000. Modeling of eicosapentaenoic acid (EPA) production from *Phaeodactylum tricorutum* cultures in tubular photobioreactors. Effects of dilution rate, tube diameter, and solar irradiance. *Biotechnology and Bioengineering*, 68 (2), 173-183.
- Gray, N. 2011. Beyond the heart and brain: emerging benefits of omega-3. In: *NutraIngredients-USA*.
- Guihéneuf, F., Fouqueray, M., Mimouni, V., Ulmann, L., Jacquette, B., Tremblin, G. 2010. Effect of UV stress on the fatty acid and lipid class composition in two marine microalgae *Pavlova lutheri* (Pavlovophyceae) and *Odontella aurita* (Bacillariophyceae). *Journal of Applied Phycology*, 22 (5), 629-638.
- Guihéneuf, F., Mimouni, V., Tremblin, G., & Ulmann, L. 2015. Light intensity regulates LC-PUFA incorporation into lipids of *Pavlova lutheri* and the final desaturase and elongase activities involved in their biosynthesis. *Journal of Agricultural and Food Chemistry*, 63 (4), 1261-1267.
- Gupta, A., Barrow, C. J., Puri, M. 2012. Omega-3 biotechnology: *Thraustochytrids* as a novel source of omega-3 oils. *Biotechnology advances*, 30(6), 1733-1745.
- Hammond, B. G., Mayhew, D. A., Naylor, M. W., Ruecker, F. A., Mast, R. W., & Sander, W. J. 2001. Safety assessment of DHA-Rich microalgae from *Schizochytrium* sp.: I. Subchronic rat feeding study. *Regulatory Toxicology and Pharmacology*, 33(2), 192-204.
- Ho, L., van Leeuwen, R., Wittelman, J. C., van Duijn, C. M., Uitterlinden, A. G., Hofman, A., de Jong, P. T., Vingerling, J. R., Klaver, C. C. 2011. Reducing the genetic risk of age-related macular degeneration with dietary antioxidants, zinc, and ω -3 fatty acids: the Rotterdam study. *Archives of ophthalmology*, 129(6), 758-766.
- Hong, W.-K., Rairakhwada, D., Seo, P.-S., Park, S.-Y., Hur, B.-K., Kim, C. H., & Seo, J.-W. 2011. Production of lipids containing high levels of docosahexaenoic acid by a newly isolated microalga, *Aurantiochytrium* sp. KRS101. *Applied Biochemistry and Biotechnology*, 164(8), 1468-1480.
- Hooper, L., Thompson, R. L., Harrison, R. A., Summerbell, C. D., Ness, A. R., Moore, H. J., Worthington, H. V., Durrington, P. N., Higgins, J. P.,

- Capps, N. E. 2006. Risks and benefits of omega 3 fats for mortality, cardiovascular disease, and cancer: systematic review. *Bmj*, 332(7544), 752-760.
- Imhoff-Kunsch, B., Stein, A. D., Martorell, R., Parra-Cabrera, S., Romieu, I., & Ramakrishnan, U. 2011. Prenatal docosahexaenoic acid supplementation and infant morbidity: randomized controlled trial. *Pediatrics*, 128(3), e505-e512.
- Khozin-Goldberg, I., Iskandarov, U., & Cohen, Z. 2011. LC-PUFA from photosynthetic microalgae: occurrence, biosynthesis, and prospects in biotechnology. *Applied Microbiology and Biotechnology*, 91(4), 905-915.
- Kiecolt-Glaser, J. K., Belury, M. A., Andridge, R., Malarkey, W. B., & Glaser, R. 2011. Omega-3 supplementation lowers inflammation and anxiety in medical students: a randomized controlled trial. *Brain, behavior, and immunity*, 25(8), 1725-1734.
- Lang, I., Hodac, L., Friedl, T., & Feussner, I. 2011. Fatty acid profiles and their distribution patterns in microalgae: a comprehensive analysis of more than 2000 strains from the SAG culture collection. *BMC plant biology*, 11(1), 124.
- Lewis, M. D., Hibbeln, J. R., Johnson, J. E., Lin, Y. H., Hyun, D. Y., & Loewke, J. D. 2011. Suicide deaths of active duty US military and omega-3 fatty acid clinical psychiatry, 72(12), 1585.
- Lozac'h, N. 1986. Extension of Rules A-1.1 and A-2.5 concerning numerical terms used in organic chemical nomenclature (Recommendations 1986). *Pure and Applied Chemistry*, 58(12), 1693-1696.
- Martins, D. A., Custodio, L., Barreira, L., Pereira, H., Ben-Hamadou, R., Varela, J., & Abu-Salah, K. M. 2013. Alternative Sources of n-3 Long-Chain Polyunsaturated Fatty Acids in Marine Microalgae. *Marine Drugs*, 11(7), 2259-2281.
- Mendes, A., Reis, A., Vasconcelos, R., Guerra, P., & da Silva, T. L. (2009). *Cryptocodinium cohnii* with emphasis on DHA production: a review. *Journal of Applied Phycology*, 21(2), 199-214.
- Newton, I. 1998. Long-chain polyunsaturated fatty acids—the new frontier in nutrition. *Lipid technol*, 10, 77-81.
- Pal, D., Khozin-Goldberg, I., Cohen, Z., & Boussiba, S. (2011). The effect of light, salinity, and nitrogen availability on lipid production by *Nannochloropsis* sp. *Applied Microbiology and Biotechnology*, 90(4), 1429-1441.
- Pase, M. P., Grima, N. A., & Sarris, J. 2011. Do long-chain n-3 fatty acids reduce arterial stiffness? A meta-analysis of randomised controlled trials. *British Journal of Nutrition*, 106(07), 974-980.
- Quilodrán, B., Hinzpeter, I., Hormazabal, E., Quiroz, A., & Shene, C. 2010. Docosahexaenoic acid (C22: 6n-3, DHA) and astaxanthin production by *Thraustochytriidae* sp. AS4-A1 a native strain with high similitude to *Ulkenia* sp.: Evaluation of liquid residues from food industry as nutrient sources. *Enzyme and Microbial Technology*, 47(1), 24-30.
- Sijtsma, L., De Swaaf, M. 2004. Biotechnological production and applications of the ω -3 polyunsaturated fatty acid docosahexaenoic acid. *Applied Microbiology and Biotechnology*, 64(2), 146-153.
- Simopoulos, A. P. (2008). The importance of the omega-6/omega-3 fatty acid ratio in cardiovascular disease and other chronic diseases. *Experimental Biology and Medicine*, 233(6), 674-688.
- Van Pelt, C. K., Huang, M. C., Tschanz, C. L., & Brenna, J. T. 1999. An octaene fatty acid, 4,7,10,13,16,19,22,25-octacosaoctaenoic acid (28 : 8n-3), found in marine oils. *Journal of Lipid Research*, 40(8), 1501-1505.
- Winwood, R. J. 2013. Recent developments in the commercial production of DHA and EPA rich oils from micro-algae. *OCL*, 20(6), D604.