# FUNCTIONAL PROPERTIES OF CAMEL MILK AND THEIR INFLUENCES ON TECHNOLOGICAL APPLICATIONS

#### Selda BULCA

University of Adnan Menderes, Faculty of Engineering, Department for Food Engineering, Aytepe/Aydin, Turkey

#### Corresponding author email: sbulca@adu.edu.tr

#### Abstract

According to the recent statistics by the Food and Agriculture Organization (FAO), the total population of camels in the world is estimated to be about 20 million, with Somalia having the largest herd worldwide. Camels are well adapted to harsh desert climates and can survive without drinking water for days. Therefore, camel (Camelus dromedarius) is of significant socio-economic importance in many arid and semi-arid parts of the world and its milk constitutes an important component of human diets in these regions.

The amounts of lysozyme, lactoferrin, lactoperoxidase and immunoglobulins were found to be greater in dromedary camel milk than bovine or buffalo milk. This property has been shown to be a disadvantage in yoghurt production. As known like yoghurt, cheese is another fermented milk product, due to the activity of these compounds the enzymatic reaction is disturbed and the gelation process of milk is prolonged. These antimicrobial agents were reported to completely lose their activity in camel milk if heat-treated at 100°C for 30 min. But there are contradictional statements about the heating intensity. Therfore in this review on these studies are focussed. In addition the chemical composition of camel milk is compared with another ruminant milks. Camel milk has lots of functional properties. These are antioxidant activity, bioactivity, anti-cancer activity, hypoallergenicity.

Key words: camel milk, lactoferrin, immunoglobulins, heat treatment, functional properties

# INTRODUCTION

There are about 18 million camels in the world (FAO, 1996) which support the survival of millions of people in arid and semi-arid areas. Meanwhile camel milk is considered one of the main components of the human diet in many parts of the world. Camels were domesticated and developed approximately 5000 years ago and throughout these years have played an integral role in the daily life of camel owners. They are distributed in Africa and Asia, where other livestock farming cannot be easily implemented (Gupta et al. 2015). Camels are very resistant animals of hunger and thirtst. Variations in the contents of camel milk may be based on to several factors such as analytical geographical area, nutrition methods. conditions, breed, lactation stage, age and number of calvings (Khaskheli et al. 2005). The quality of camel milk and meat, since it contains both valuable essential nutrients, has acquired an important place in human nutrition (Adel et al. 2009). According to the recent statistics by the Food and Agriculture Organization (FAO, 2008), the total population of camels in the world is estimated to be about 20 million, with Somalia having the largest herd worldwide (FAO, 2008).

Also according to FAO data the production of camel milk is 5.3 million/liter in the world. At the present time, depending on the camel cultivation camel milk production is also becoming increasingly common.

For this reason, the number of scientific research on camel milk have increased in recent years. They are mainly distributed in African and Asian arid and semi-arid areas, where other livestock farming cannot be easily applied (Gupta *et al.* 2015). Today, camels and their products have been using by humans for transport, traction power, milk, meat, fiber (wool and hair). At the same time, it is used as a raw material for textile industry.

#### **Chemical Composition of Camel Milk**

The camel has the ability to produce more milk for a longer period of time in arid zones and dry lands (an environment of extreme temperature, drought, and lack of pasture) than in other domestic livestock species (Yagil and Etzion, 1980).

Geographical root and seasonal variations are factors which influence most changes in composition of camel milk. Camel milk contains 2.9 to 5.5% fat, 2.5 to 4.5% protein, 2.9 to 5.8% lactose, 0.35 to 0.90% ash, 86.3 to 88.5% water, and 8.9 to 14.3% solid-non-fat (SNF) (Khan and İkbal, 2001). Camel milk has similar protein content, lower lactose content (Elamin & Wilcox, 1992), and greater total cholesterol (Gorban and Izzeldin, 1999) compared with cow's milk. Camel milk has greater contents of vitamin C (Mehaia, 1994), ash, and sodium, potassium, phosphorus, zinc, iron and manganese (Gorban and Izzeldin, 1997) than cow's milk.

Seasonal variations also play a significant role in the composition of camel milk, also with camels of the same type and from the same district (Bakheit et al. 2008).

According to other research related to compositional, technological and nutritional aspects of dromedary camel milk the average values of camel milk composition reported from 1980 to 2009 are as follows: protein 3.1%; fat 3.5%; lactose 4.4%; ash 0.79% and total solids 11.9% (Adel et al. 2009). Rates of milk components are based on various types of animals.

Camel's milk is a good source of various vitamins and minerals and it has several medicinal and therapeutic effects and good antibacterial and antiviral properties (Yagil & Etzion, 1980; Balouiri et al., 2016). Some studies showed that camel's milk is an excellent source of components that are involved in some biological activities, such as defence against free radicals and reactive oxygen species. The world's total population of camels was reported to be twenty-two million in 2010 (FAO, 2012) that could produce about 300 million litres milk representing 0.2% of world's total produced milk in 2010 (IDF, 2010).

The amounts of lysozyme, lactoferrin and immunoglobulins were found to be greater in dromedary camel milk than bovine or buffalo milk (Benkerroum, 2008; El-Agamy et al., 2000; Kappeler et al., 1999; Konuspayeva et al., 2007). This property has been shown to be a disadvantage in yoghurt production.

The growth of yoghurt culture in camel milk is delayed due to the presence of lysozyme (Abu-Tarboush, 1996; Jumah, Shaker, & Abu-Jadayil, 2001) which prolongs the gelation process (Jumah et al., 2001).

These antimicrobial agents were reported to completely lose their activity in camel milk if heat-treated at 100°C for 30 min (El-Agamy, 2000).

According to another observations and experiments unlike cow milk, it was found that camel milk can be preserved for a longer time at 30°C and most importantly the camel milk can be kept at 4°C for more that three months without any appearing change (Yagil, 1985).

The ability of camel milk to inhibit growth of pathogenic bacteria and its relation to whey lysozyme has been showed by Barbour et al. (1984).

At the same time, camel milk is higher in  $\alpha$ lactalbumin, as it is in human milk compared with cow milk.

Unpublished commercial data reported that some infant formula contains high level of  $\alpha$ -lactalbumin in changing to breast feed milk.

# Antimicrobial factors of camel and human milk

As shown in Table 1 camel milk is richer in immunoglobulins than human milk. However, its contents of lactoferrin and lysozyme were very low. El-Agamy and Nawar (2000) found that camel milk is contain 1.64 mg/ml of immunoglobulin G versus 0.67, 0.63, 0.70, 0.55 and 0.86 for cow, buffalo, goat, sheep and human milk, respectively.

A comparative study of lysozyme concentration in milk of different species (El-Agamy et al., 1997) showed that camel milk contained significantly higher content of lysozyme than cow, buffalo, sheep and goat but very low content as compared to lysozyme content of human, mare and donkey milks.

The same study showed that camel milk contained also signi ficantly higher level of lactoferrin (0.22 mg/ml) than cow, buffalo, sheep and goat but very low compare with that of human milk.

Antimicrobial factor	Camel milk	Human milk
Mean values $\pm$ SD		
Immunoglobulins (mg/ml)	$1.54\pm0.032$	$1.14\pm0.055$
Lactoferrin(mg/ml)	$0.24\pm0.035$	$1.95\pm0.050$
Lysozyme(mg/ml)	$0.06\pm0.02$	$0.65\pm0.045$

### Table 1. Antimicrobial factors in camel and human milks (El-Agamy et al., 1997)

# Nutritional properties of camel and human milks

Milk of all mammals contains the same principal components, namely water, proteins, fats, carbohydrates, vitamins and minerals, but their content varies widely between ruminant and nonruminant milk. Especially, camel milk contains all essential nutrients as cow milk (El-Agamy et al., 1998). Many components in bovine colostrum and milk exhibit specific biological activity in addition to their established nutritional values. During the past two decades, interest in these beneficial physiological effects and the possibility to utilise the components from milk have increased.

Even between various (non-) ruminants and within a same species the milk composition may differ considerably, given the influence of genetic factors (not only at species but also at breed level), physiological factors (e.g. lactation stage, milking interval), nutritional factors (e.g. feed energy value and composition) and environmental conditions (e.g. location, season). The values should therefore not be viewed as absolute but rather as indicative for the concentration range of milk components. Moreover, methodological differences regarding data collection between consulted papers may contribute to the spread of the presented values.

# Therapeutic properties of Camel milk

According to studies, the production of camel milk has significantly increased during the last few years with now pasteurized fresh camel milk in the supermarket. Firstly, camel milk is supposed to have medicinal properties (El-Agamy et al., 1992). In studies camel milk is used jaundice, asthma, in the treatment of various diseases such as tuberculosis and it has been found to be helpful. In addition to this column, cancer, diabetes, hypertension was identified that help to treat their patients (Hossam, 2015). Nowadays, there is a general need to start a number of camel milk based functional products to the commercial markets due to increasing demand in recent years (Al haj *et al.* 2010). These products have to be clinically proven and scientifically evident supported (Ghosh, 2009). Camel milk has lots of functional properties.

These are antioxidant activity, bioactivity, anticancer activity, hypoallergenicity activity (Habib et al., 2013). It is also known that the camel milk has a therapeutic potential against many diseases including cancer. In addition it has long been utilized for its benefit in broad range of diseases like Insulin Dependent Diabetes Mellitus (IDDM) (Agrawal et al., 2002; Agrawal et al., 2003; Agrawal et al., 2005), infant diarrhea (Yagil, 2013), hepatitis (El-Fakharany et al., 2008), allergy, lactose intolerance (El-Agamy et al., 2009; Konuspayeva et al., 2009; Cardoso et al. 2010). It contains extraordinarily high levels of insulin like molecule (Agrawal et al., 2002; 2003; 2005).

Camel milk is emerging as a potent therapeutic alternative which can help in reducing insulin doses in diabetic patients. It's well established role in management of Diabetes has rendered it the title of "white gold of desert". Epidemiological surveys strongly indicate low prevalence of diabetes in communities consuming camel milk. (Agrawal et al., 2013).

# Composition of Camel milk colostrum

Colostrum is a complex fluid rich in nutrients and is also characterised by its high level of bioactive components, e.g. immunoglobulins especially IgG1, growth (Igs), factors. especially insulin-like growth factor-1 (IGF-1), transforming growth factor beta-2 (TGF-b2) and grow th hormone (GH) as well as lactoferrin, lysozyme andlacto peroxidase (Butler, 1994; Pakkanen, 1998; Regester, Smithers, Mitchell, McIntosh, & Dionysius, 1997; Reiter, 1985). Camel colostrum differs in composition from regular milk in that it has a high content of whey proteins, mainly immunoglobulins G (IgG), providing the newborn with immunity.

Camel colostral IgG consists of three main subclasses, namely IgG1, IgG2, and IgG3 (Azwai et al., 1996) the two latter sub-classes are devoid of light chains and have a molecular mass of 42 and 45 kDa, respectively (Hamers-Casterman et al., 1993). It has been reported that these heavy-chain antibodies interfere with several biological processes and may make it a good candidate for human therapy (Holt et al., 2003). To current knowledge, no information is available regarding the variation in IgG and other major whey proteins in camel colostrum and milk during the first week of lactation.

### Antibacterial activity of Camel milk

Camel milk is reported to have an antimicrobial effect against Gram positive and Gram negative bacteria, including Escherichia coli, Listeria monocytogenes, Staphylococcus aureus and Salmonella typhimurium (Benkerroum et al., 2004; El-Agamy and Khatab, 1992). This inhibitory activity was attributed to the presence of antimicrobial substances in camel milk, including lysozyme, hydrogen peroxide, Lactoperoxidase lactoferrin, and immunoglobulins (El-Agamy and Khatab, 1992). Lactoperoxidase (LPO) is a suitable enzymatic indicator of correct pasteurisation of camel milk and its products are heat-treated at 75 degrees C for 15 seconds (Wernerv et al., 2013).

The inhibitory action of camel milk against *L.* monocytogenes, *S. aureus* and *E. coli* might be attributed to the presence of lactoperoxidase, hydrogen peroxide and lysozyme respectively (Benkerroum et al., 2004). The growth of *Salmonella Typhimurium* was inhibited by lactoferrin in camel milk through binding iron and making it unavailable for its growth (El-Agamy and Khatab, 1992; Ochoa & Cleary, 2009).

### Antibacterial activity of Transferrin

Transferrin (TF) is a monomeric glycoprotein of 679 amino acids, with a relative molecular weight of approximately 80 kDa. Transferrin exists mainly in the serum and interstitial compartments of vertebrates and some invertebrates (Baker and Lindley, 1992; Retzer et al., 1996). Transferrin is found at a much lower concentration in human milk (< 50  $\mu$ g/mL) in comparison with bovine milk (20 to 200  $\mu$ g/mL) (Schanbacher et al., 1993). The principle physiological function of TF in mammals is to transport ferric irons from sites of absorption to sites of utilization. Transferrin transports iron from the biological fluids into the cytoplasm via plasma membrane by receptor-mediated endocytosis (Pakdman and Chahine, 1997). Transferrin interacts with specific receptors present in variable amounts on target cells. Important target cells include the liver, bone marrow and muscle.

Antibacterial activity of Lactoferrin Lactoferrin is a mammalian cationic ironbinding glycoprotein belonging to the transferrin family, which was discovered 70 years ago, and isolated simultaneously from human and bovine milks in 1960. Lactoferrin is present in the majority of external secretions and mucosal surfaces, milk being its main source. Lactoferrin binds two atoms of iron and due to this capacity several functions have been attributed to it. such as antibacterial. antioxidant. antitumoral and immunomodulatory (Sanchez et al., 1992).

It is widely distributed in all biological fluids and is also expressed by immune cells, which release it under stimulation by pathogens. Lactoferrin is a multi-functional protein with many beneficial properties, which makes it a functional food for a number of product, commercial and clinical applications (Adlerova et al., 2008). Lactoferrin is a glycoprotein with a molecular weight of about 80 kDa, which shows high affinity for iron. The molecular structure and amino acid sequence of human lactoferrin were discovered in 1984.

Almost all bacteria require iron for their growth; therefore LF devoid of iron is capable of preventing its utilization by some bacteria (Orsi, 2004). A large number of studies have demonstrated the bacteriostatic and bactericidal effect of LF, against a wide range of Grampositive and Gram-negative bacteria (Farnaud and Evans, 2003). However, other mechanisms besides iron holding can be involved in the antibacterial activity of LF, such as blocking microbial metabolism of carbohydrates or destabilizing the bacterial cell wall (Sanchez et al., 1992).

### Antibacterial activity of Lyzozyme

Lysozyme (EC 3.2.1.17; muramidase) is a single polypeptide chain consisting of 129

amino acids, in which lysine is the N-end amino acid and leucine is the C-end one. It is a globular basic protein characterized bv molecular weight of 14.3 kDa and cross-linked by four disulfide bonds (Masschalck et al., 2002; Cegielska et al., 2008). It is an important antimicrobial agent in milk, which kills bacteria by cleaving the  $\beta$ -1,4-glycosidic bond between C-1 of N-acetvl muramic acid and C-4 of Nglucosamine residues acetvl of the peptidoglycan in the bacterial cell wall (Zhao, et al., 2011; Li et al., 2011). Lysozyme appears to inhibit not only bacteria where the peptidoglycan layer is a major component of their cell-wall, but also viruses and eukaryotic microorganisms devoid of а typical peptidoglycan layer, suggesting that it acts by other mechanisms of action than the hydrolytic activity (Benkerroum, 2008)

### Antibacterial activity of Immunoglobulins

Immunoglobulins in milk immediately brings to mind the relationship between mother's milk, transfer of passive immunity from mother to neonate, and the immature immune system of the neonate. Research in this field dates back to the late nineteenth century, however for many centuries herdsmen have capitalized on the linkage between maternal immune status immunological the protection and and development of the neonate (Butler, Kehrli, 2005; Wheeler et al., 2007). Immunoglobulins in mammary secretions come from several sources and represent a history of the antigen exposure of the mother and the response of her immune system. Immunoglobulins are transported through the mammary epithelial cells by receptor-mediated mechanisms and transferred out of the mammary gland by milk ejection during suckling. The immunoglobulins then enter the environment of the gastrointestinal tract of the neonate. Although that environment is primarily geared toward digestion to gain nutritional benefit, the immunoglobulins remain sufficiently stable to provide protective benefits for the neonate, either through uptake into the vascular system in the newborn of some species or through immunological function in the gastrointestinal tract. The immunoglobulins found in milk and the transfer of passive immunity from mother

to neonate have been reviewed by many authors.

# Antifungal activity of camel milk components

Regarding the antifungal activity of lactoferrin, the first observation which can be made is that the great majority of research has been carried out on Candida, well known as one of the most dangerous opportunistic pathogens. As for bacteria, the anti-Candida activity of lactoferrin was initially considered as re lated to its ability to bind and sequester environmental iron. But in addition to the iron-chelating activity, a direct interaction between lactoferrin and Candida cells was demonstrated in our Department by Valenti et al. (1986).

# Antiviral activity of camel milk components

In a few cases it is reported that lactoferrin failed to prevent virus infection. On the contrary, a long list of virus has been found to be sensitive to the inhibiting action of lactoferrin. This list includes several enveloped viruses such herpes simplex virus 1 and 2 (Hasegawa et al. 1994). human cytomegalovirus (Hasegawa et al. 1994), human immunodeficiency virus (Harmsen et al. 1995), hepatitis B virus (Hara et al. 2002), hepatitis C virus (Ikeda et al. 1998), respiratory syncytial virus (Grover et al. 1997), hanta virüs (Murphy et al. 2000) and four naked viruses: rotavirus (Superti et al. 1997), poliovirus (Marchetti et al. 1999), adenovirus (Arnold et al. 2002) and enterovirus 71 (Lin et al. 2002).

# Evaluation of camel's milk from technological aspects

The absence of  $\beta$ -LG might explain some of the differences observed between camel and cow milk regarding technological properties such as thermal stability during drying, heat induced aggregation and adherence to heating surfaces (fouling properties) as well as the thin consistency found in fermented camel milk (Merin et al. 2001; El-Agamy, 2007; El-Hatmi et al. 2007; Laleye et al. 2008).

A detrimental effect of heating on the beneficial health effects of milk, the most frequently cited arguments of raw milk advocates are a reduced susceptibility to allergies, a higher nutritional quality and a better taste. However, the consumption of raw milk poses a realistic microbiological risk for the consumer. The presence of foodborne pathogens has been demonstrated in many surveys and foodborne infections have been repeatedly reported for Campylobacter. Salmonella spp. and human pathogenic verocytotoxin-producing Escherichia coli after raw milk consumption (Claeys et al., 2013; O'Mahony, Fanning and Whyte, 2009: Robinson, Scheftel, & Smith, 2014; Verraes et al., 2014).

Whey proteins of bovine milk are less resistant to heat denaturation compared to those of buffalo milk, which in turn are less heat resistant than camel whey proteins (El-Agamy, 2000). Even though camel whey proteins have a higher heat stability than bovine whey proteins at temperatures between 63 and 90°C (Farah, 1986), bovine milk coagulates much slower at higher temperatures. This could be related to the absence or very low levels of  $\beta$ -lg and  $\kappa$ -casein in camel milk (Farah & Atkins, 1992) as milk is more resistant to heat when it is characterized by a molar  $\beta$ -lg to  $\kappa$ -casein ratio close to 1 (Barlowska, Szwajowska, Litwinczuk, Król, 2011).

In another study is shown that the heat stability of camel milk was relatively lower at high temperature treatments. Heat coagulation time (HCT) in the range 100-130 degrees C was too short (< 2 min). Camel milk heat preservation can be done only by pasteurisation. After LTLT pasteurisation, counts of aerobic total and psychrotrophic bacteria were significantly (p < 0.05) reduced and coliforms were not detected (Kouniba et al., 2005).

There are some investigations applied the inactivity of enzymes, which have from the technological sides important. In this case the activity of alkaline phosphatase (ALP). gamma-glutamyltransferase (GGT), lactoperoxidase (LPO), lipase (LIP) and leucine arylamidase (LAP) in raw and pasteurised camel milk was studied, in order, to find a heat treatment indicator suitable to verify an effective pasteurisation. LAP activity in raw camel milk is too low and the data variation is too high for serving as a marker. The LPO results look promising. The enzyme activity in raw camel milk is high and the respective value in pasteurised milk is predominantly below the detection limit of the method.

## CONCLUSIONS

The production of camel milk is gradually increasing due to an increased interest by consumers in recent years. Camel milk was found to be different in some aspects from milk of other animal species, such as bovine milk. Use of camel milk is widespread not only during production of different kinds of milk products but also as cure material to heal different kinds of diseases such as cancer, diabetes, hypertension, autism dropsy, jaundice, tuberculosis, asthma.

Except from the therapeutic properties the use of camel milk is investigated in different area. Due to functional properties of some camel milk components such as lactoferrin, lyzozyme and immunoglobulins the camel milk is longer storable than other kinds of ruminants milk. Some of functional properties are called such as antibacterial, antiviral, antifungal, antiallergic exc.

Camel milk is known as an alternative milk source and is widespread in many countries. The production of milk products such as yoghurt, chees, ice-cream, pasteurised milk especially in Somalia and Sudan.

### REFERENCES

- Abu-Tarboush HM., 1996. Comparison of Associative Growth and Proteolytic Activity of Yogurt Starters in Whole Milk from Camels and Cows. Journal of Dairy Science, 79: 366-371
- Adel O., Niamh H., Oruna-Concha MJ., 2009. Quantification of major camel milk proteins by capillary electrophoresis. International Dairy Journal, 58: 31-35
- Adlerova L., Bartoskova A., Faldyna M., 2008. Lactoferrin: a review. Veterinarni Medicina, 53: 457-468
- Agrawal RP., Beniwal R., Kochar DK., Tuteja FC., Ghorui SK., Sharma S., 2005. Camel milk as an adjunct to insulin therapy improves long-term glycemic control and reduction in doses of insulin in patients with type-1 diabetes A 1 year randomized controlled trial. Diabetes Res. Clin. Practice, 68: 176-177
- Agrawal RP., Swami SC., Beniwal R., Kochar DK., Sahani MS., 2003. Effect of camel milk on glycemic control, lipid profile and diabetes quality of life in type 1 diabetes: A randomized prospective controlled cross over study. Indian J. Anim. Sci. 73: 1105-1110.

- Agrawal RP., Swami SC., Beniwal RD., Kochar K., Kothari RP., 2002. Effect of camel milk on glycemic control, risk factors and diabetes quality of life in type-1 diabetes: A randomized prospective controlled study. Int. Diabetes Dev. Ctries. 22: 70-74
- Al Haj O., Al Kanhal A., Hamad A., 2010. Compositional, technological and nutritional aspects of dromedary camel milk. International Dairy Journal, 20: 811-821
- Arnold RR., Cole MF., Mcghee JR., 1977. A bactericidal effect for human lactoferrin. Science 197: 263–265
- Azwai SM, Carter SD., Woldehiwet Z., 1996. Immunoglobulins of Camel (*Camelus dromedarius*). Colostrum. J. Comp. Path., 114: 273-282
- Baker EN., Lindley PF., 1992. New Perspectives on the Structure and Function of Transferrins. Journal Inorg. Biochem, 47: 147–160
- Bakheit SA., Majid AMA., Nikhala AM., 2008. Camels (*Camelus dromedarius*) under pastoral systems in North Kordofan, Sudan: seasonal and parity effects on milk composition. Journal of Camelid Science, 1: 32-36
- Balouiri M., Sadiki M., Ibnsouda SK., 2016. Methods for in vitro evaluation antimicrobial activity: A review. Journal of Pharmaceutical Analysis, 6:71–79
- Barbour EK., Nabbut NH., Frerichs WM., Alnakhli HM., 1984. Inhibition of pathogenic bacteria by camels milk-Relation to whey lysozyme and stage of lactation 47: 838-840
- Barlowska J., Szwajowska M., Litwinczuk Z., Król J., 2011. Nutritional value and technological suitability of milk from various animal species used for dairy production. Comprehensive Reviews in Food Science and Food Safety, 10: 291-302
- Benkerroum N., 2008. Antimicrobial activity of lysozyme with special relevance to milk. African Journal of Biotechnology, 25: 4856–4867
- Benkerroum N., Mekkoui M., Bennani N., Hidane K., 2004. Antimicrobial activity of camel's milk against pathogenic strains of *Escherichia coli* and *Listeria monocytogenes*. International Journal of Dairy Technology, 57: 39-43
- Butler JE., 1994. Passive immunity and immunoglobulin diversity. In Proceedings of the IDF seminar: Indigenous antimicrobial agents of milk-recent developments, Brussels International Dairy Federation 14–50
- Butler JE.; Kehrli ME., 2005. Jr. Immunoglobulins and immunocytes in the mammary gland and its secretions. In Mestecky J., Lamm M., Strober W., Bienenstock J., McGhee JR., Mayer L., 3rd ed *Mucosal Immunology*, Elsevier Academic Press: Burlington MA USA, Volume 2, pp. 1764–1793.
- Cardoso RA., R. Santos MDB., Cardoso CRA., Carvalho MO., 2010. Consumption of camel's milk by patients intolerant to lactose. A preliminary study. Rev. Alergia México, 57: 26-32
- Cegielska RR., Lesnierowski G., Kijowski J., 2008. Properties and application of egg white lysozyme and its modified preparations-A review. Polish Journal of Food and Nutrition Sciences, 58 (1): 5–10
- Claeys WL., Cardoen S., Daube G., De Block J., Dewettinck K., Dierick K., 2013. Raw or heated cow

milk consumption: review of risks and benefits. Food Control, 31: 251-262

- El Hatmi H., Giradet JM., Gaillard JL., Yahyaoui MH., Attia H., 2007. Characterization of whey proteins of camel (Camelus dromedarius) milk and colostrums. Small Rum. Res. 70: 267-271
- El-Agamy EI., 2000. Effect of heat treatment on camel milk proteins with respect to antimicrobial factors: a comparison with cows' and buffalo milk proteins. Food Chemistry, 68: 227-232
- El-Agamy EI., 2007. The challenge of cow milk protein allergy. Small Ruminant Research, 68: 64-72
- El-Agamy EI., Abou-Shloue ZI., Abdel-Kader YI., 1997. A comparative study of milk proteins from different species. II. Electrophoretic patterns, molecular characterization, amino acid composition and immunological relationships. Third Alexandria Conference on Food Science and Technology, Alexandria, Egypt, March, 1-3
- El-Agamy EI., Abou-Shloue ZI., Abdel-Kader YI., 1998. Gel electrophoresis of proteins, physicochemical characterization and vitamin C content of milk of different species. Alexandria J. Agric. Res., 43(2): 57-70.
- El-Agamy El., Abou-Shloue, F., Abdel-Kader A., 2000. Effect of heat treatment on camel milk proteins with respect to antimicrobial factors: a comparison with cows' and buffalo milk proteins. Food Chemistry 68: 227–32
- El-Agamy EI., Khatab AA., 1992. Physicochemical and microbiological characteristics of Egyptian human milk. Alexandria J. Agric. Res., 37(2): 115-126
- El-Agamy EI., Nawar M., 2000. Nutritive and immunological values of camel milk, A comparative study with milk of other species. In: Proc. 2nd International Camelid Conference: Agroecons. Camelid Farm. Almaty, Kazakhstan, 8-12 Sept
- El-Agamy El., Nawar, MS., Shamsia S., Awad and GF., Haenlein W., 2009. Are camel milk proteins convenient to the nutrition of cow milk allergic children? Small Rumin. Res., 82: 1-6
- El-Agamy El., Ruppanner R., Ismail A., Champagne CP., Assaf R., 1996. Purification and characterization of lactoferrin, lactoperoxidase, lysozyme and immunoglobulins from camel's milk. Inter. Dairy Journal, 6: 129-145.
- Elamin FM., Wilcox CJ., 1992. Milk composition of Majaheim camels. Journal of Dairy Science, 75: 3155–3157
- El-Fakharany EM., Tabll A., El-Wahab AA., Haroun MB., El-Rashdy RM., 2008. Potential Activity of Camel Milk-Amylase and Lactoferrin against Hepatitis C Virus Infectivity in HepG2 and Lymphocytes. Hepatitis Monthly, 8(2): 101-109
- FAO 1996. Statistics year-book, FAO, Rome.
- FAO 2008. Camel milk, Retrieved from Farah Z., Ruegg MW., The size distribution of casein micelles in camel milk. Food Microstructure 8: 211–212
- FAO 2012. http://www.fao.org/. 02.05.2012
- Farah Z., 1986. Effect of heat treatment on whey proteins of camel milk. Milchwissenschaft, 41: 763-765

- Farah Z., Atkins D., 1992. Heat coagulation of camel milk. Journal of Dairy Research, 59: 229-231
- Farnaud S., Evans RW., 2003. Lactoferrin-a multifunctional protein with antimicrobial properties. Molecular Immunology, 40: 395-405
- Ghosh D., 2009. Future perspectives of nutrigeonomics foods: Benefits vs risks. Indian Journal of Biochemistry & Biophysics, 46: 31-36
- Gorban AMS., Izzeldin OM., 1997. Mineral content of camel milk and colostrum. Journal of Dairy Research, 64: 471–474
- Gorban AMS., Izzeldin OM., 1999. Study on cholesteryl ester fatty acids in camel and cow milk lipid. International Journal of Food Science and Technology, 34: 229–234
- Grover M., Giouzeppos O., Schnagel RD., May JT., 1997. Effect of human milk prostaglandins and lactoferrin on respiratory syncityal virus and rotavirus. Acta Paediatr 86: 315–316
- Gupta BM., Mueen KK., Ritu A., Rishi T., 2015. World camel research: a scientometric assessment. Scientometrics, 102: 957–975
- Habib HH., Ibrahim WH., Schneider-Stock R., Hassan, MH., 2013. Camel milk lactoferrin reduces the proliferation of colorectal cancer cells and exerts antioxidant and DNA damage inhibitory activities. Food Chemistry, 141: 148–152
- Hamers-Casterman C., Atarhouch T., Muyldermans S., Robinson G., Hamers C., Songa EB., Bendahman B., Hamers R., 1993. Naturally occurring antibodies devoid of light chains. Nature, 363, 446–448.
- Hara K., Ikeda M., Saito S., 2002. Lactoferrin inhibits hepatitis B virus infection in cultured human hepatocytes. Hepatol Res 24: 228-236
- Harmsen MC., Swart PJ., de Bethune MP., 1995 Antiviral effects of plasma and milk proteins: lactoferrin shows potent activity against both human immunodeficiency virus and human cytomegalovirus replication in vitro. J Infect Dis 172: 380–388
- Hasegawa K., Motsuchi W., Tanaka S., Dosako S., 1994. Inhibition with lactoferrin of in vitro infection with human herpes virus. Jpn JMed Sci Biol 47: 73–85
- Holt LJ., Herring C., Jespers LS., Woolven BP., Tomlinson IM., 2003. Domain antibodies: proteins for therapy. Trends Biotech. 21: 484–490
- Hossam E., Bahaa AS., Iftekhar H., Jameel Al-T., Metwalli A., Alhazza I., 2015. Camel milk peptide improves wound healing in diabetic rats by orchestrating the redox status and immune response. Lipids in Health and Disease, 14:132-135
- IDF 2010 International Dairy Federation. A common carbon footprint or dairy, The IDF guide to standard lifecycle assessment methodology for the dairy industry. International Dairy Federation
- Ikeda M., Sugiyama K., Tanaka T., 1998 Lactoferrin markedly inhibits hepatitis C virus infection in cultured human hepatocytes. Biochem Biophys Res Commun 245: 549–553
- Jumah RY., Shaker RR., Abu-Jadayil B., 2001. Effect of milk source on the rheological properties of yoghurt during the gelation process. International Journal of Dairy Technology, 54: 89-93

- Kappeler SR., Ackermann M., Farah Z., Puhan Z., 1999. Sequence analysis of camel (*Camelus dromedarius*) lactoferrin. International Dairy Journal, 9: 481-486
- Khan BB., Iqbal A., 2001. Production and composition of camel milk. Pakistan Journal of Agricultural Sciences, 38: 64–67
- Khaskheli M., Arain MA., Chaudhry S., Soomro AH., Qureshi TA., 2005. Physico-chemical quality of camel milk. Journal of Agriculture and Social Sciences, 2: 164-166
- Konuspayeva G., Faye B., Loiseau G., 2007. Lactoferrin and immunoglobulin contents in camel's milk (Camelus bactrianus, Camelus dromedaries, and hybrids) from Kazakhstan. Journal of Dairy Science, 90: 38-46
- Konuspayeva G., Faye B., Loiseau G., 2009. The composition of camel milk: A meta-analysis of the literature data. J. Food Comp. Anal. 22: 95-101
- Kouniba A., Berrada M., Zahar M., 2005. Composition and heat stability of moroccan camel milk, Journal of Camel practice and research, 12: 105-110
- Laleye LC., Jobe B., Wasesa AAH., 2008. Comparative Study on Heat Stability and Functionality of Camel and Bovine Milk Whey Proteins, Journal of Dairy Science, 91: 4527–4534
- Li D., Zhang T., Xu C., Ji B., 2011. Effect of pH on the interaction of baicalein with lysozyme by spectroscopic approaches. Journal of Photochemistry and Photobiology, 104: 414–424.
- Lin TY., Chu C., Chiu CH., 2002. Lactoferrin inhibits enterovirus 71 infection of human embryonal rhabdomyosarcoma cells in vitro. J Infect Dis 186: 1161–1164
- Marchetti M., Superti F., Ammendolia MG., Rossi O., Valenti P., Seganti L., 1999. Inhibition of poliovirus type 1 infection by iron-, manganese- and zincsaturated lactoferrin. Med Microbiol Immunol 187: 199–204
- Masschalck B., Deckers D., Michiels M., 2002. Lytic and nonlytic mechanism of inactivation of grampositive bacteria by lysozyme under atmospheric and hydrostatic pressure. Journal of Food Protection, 65: 916–923.
- Mehaia MA., 1994. Vitamin-C and riboflavin content in camels milk-effects of heat treatments, Food Chemistry, 50: 153–155
- Merin U., Bernstein S., Bloch-Damti A., Yagil R., van Creveld C., Lindner P., Gollop N., 2001. A comparative study of milk serum proteins in camel (*Camelus dromedarius*) and bovine colostrum, Livestock Production Science, 67: 297–301
- Murphy ME., Kariwa H., Mizutani T., Yoshimatsu K., Arikawa J., Takashima I.. 2000 In vitro antiviral activity of lactoferrin and ribavirin upon hantavirus. Arch Virol 145: 1571–1582
- O'Mahony M., Fanning S., Whyte P., 2009. The safety of raw liquid milk. In Tamine AY., (Ed.), Milk processing and quality management. West Sussex (UK): Blackwell Publishing Ltd./John Wiley & Sons Ltd. 139-167
- Ochoa TJ., Cleary TG., 2009. Effect of lactoferrin on enteric pathogens. Biochimie, 91: 30-34

Orsi N., 2004. The antimicrobial activity of lactoferrin: current status and perspectives. Biometals, 17, 189-196

- Pakdman R., El Age Chahine J., 1997. Transferrin: Interaction of Lactoferrin with Hydrogen Carbonate. Eur. J. Biochem. 249: 149–155
- Pakkanen R., Aalto J., 1997. Growth factors and antimic robial factors of bovine colostrum. International Dairy Journal, 7: 285–297.
- Regester GO., Smithers GW., Mitchell IR., McIntosh GH., Dionysius DA., 1997. Bioactive factors in milk: Natural and induced. In Welch RAS., Burns DJW., Davis SR., Popay AI., Prosser CG., (Eds.), Milk composition, production and biotechnology, New York: CAB International, 119–132
- Reiter B., 1985. Protective proteins in milk—biological significance and exploitat ion. Bulletin of IDF, 191: 1–35.
- Retzer MD., Kabani A., Button LL., Yu Y., Schryvers AB., 1996. Production and Characterization of Chimeric Transferrins for the Determination of the Binding Domains for Bacterial Transferrin Receptors. J. Biol. Chem., 271: 1161–1173
- Robinson A., Trisha J., Scheftel C., Joni M., Smith, KE., (2014). Raw milk consuption among patients with Non outbreak related Enteric Infections, Emerging infectious diseases, 20: 38-44
- Sanchez L., Calvo M., Brock JH., 1992. Biological role of lactoferrin. Archives of Disease Childhood, 67: 657-661
- Schanbacher FL., Goodman REG., Talhouk RS., 1993. Bovine Mammary Lactoferrin: Implications from Messenger Ribonucleic Acid (mRNA) Sequence and Regulation Contrary to Other Milk Proteins. J. Dairy Sci., 76: 3812–3831

- Superti F., Ammendolia MG., Valenti P., Seganti L., 1997. Antirotaviral activity of milk proteins: lactoferrin prevents rotavirus infection in the enterocyte-like cell line HT29. Med Microbiol Immunol 186: 83–91
- Valenti P., Greco R., Pitari G., 1999. Apoptosis of Caco-2 intestinal cells invaded by *Listeria monocytogenes*: protective effect of lactoferrin. Exp Cell Res 250: 197–202
- Verraes C., Claeys W., Cardoen S., Daube., G., De Zutter L., Imberechts H., Dierick K., Herman L., 2014. A review of the microbiological hazards of raw milk from animal species other than cows. International Dairy Journal 39: 121-130
- Wernery U., Wernery R., Masko O., 2013. Lactoperoxidase: A suitable enzymatic marker of camel milk pasteurisation, Journal of camel practice and research, 20: 35-38
- Wheeler TT., Hodgkinson AJ., Prosser CG., Davis SR., 2007. Immune components of colostrum and milk-A historical perspective. J. Mammary Gland Biol. Neoplasia, 12: 237–247.
- Yagil R., 1985. The Desert Camel: Comparative Physiology. Comparative Animal Nutrition. FAO Animal production and healt paper 26, Rome Vol 5. Basel: Karger Ag
- Yagil R., Etzion Z., 1980. Effect of drought on the quality of camel milk. Journal of Dairy Research, 47: 159-166
- Yagil Y., 2013. Camel milk and its unique anti-diarrheal properties. Israel Med. Assoc. J. 15: 35-36
- Zhao L., Sun J-S., Sun L., 2011. The g-type lysozyme of scophthalmus maximus has a broad substrate spectrum and is involved in immune response against bacterial infection. Fish and Shelfish Immunology, 30: 630–637