MICROBIAL BIOSTIMULANTS INCREASE BIOACTIVE COMPOUNDS IN MEDICINAL PLANTS - A REVIEW

Lavinia-Diana-Nicoleta BARBU^{1, 2}, Oana-Alina BOIU-SICUIA^{*1, 2}, Călina Petruța CORNEA¹, Narcisa BĂBEANU¹

¹University of Agronomic Sciences and Veterinary Medicine of Bucharest, Faculty of Biotechnology, 59 Marasti Blvd., District 1, Bucharest, Romania ²Research-Development Institute for Plant Protection, 8 Ion Ionescu de la Brad Blvd., District 1, Bucharest, Romania

Corresponding author email: sicuia oana@yahoo.com

Abstract

Functional foods are gaining popularity among consumers because they contain biologically active compounds involved in human health and wellness. Although some plant species are already known for their beneficial properties if properly consumed, certain cultivation technologies can increase their content of bioactive compounds. For this purpose, plant probiotics could be applied as biostimulants, which are approved in organic agriculture. Plant probiotics are beneficial microorganisms that can be found in the rhizosphere, or as endophytes. They can be applied as single strains or as consortia. These beneficial microorganisms could increase the production of biologic active compounds in their host plants, such as: flavonoids, organic acids, phenols, tannins, and vitamins. This review focuses on various plant probiotics and their role in the bioactive compounds synthesis in their hosts. Among plant probiotics mediating bioactive compounds synthesis in plants are mentioned Azospirillum, Azotobacter, Bacillus, Phyllobacterium, Pseudomonas, Rhizobium and several other genera of plant beneficial microorganisms.

Key words: plant probiotics, beneficial bacteria, endophytes, medicinal bioactive compounds, functional foods.

INTRODUCTION

Functional foods, nutraceuticals, and designer are edible products foods containing supplements which complete diets and improve consumer's health. The functional foods, beside their nutritional value, provide unexpected for benefits the consumer. comparison, designer foods are fortified with promoting ingredients, health nutraceuticals are derived from food sources and used as additional health improvers to complete the basic nutritional value found in foods. All these became increasingly popular in our times, due to the fact they bring additional fiber, vitamins, minerals, biologic active compounds, prebiotics and probiotics in our diets. Nowadays, some spontaneous herbs also acquired unexpected credentials. Less popular plant species, such as amaranth (Amaranthus spp.), sorrel (Rumex spp.), purslane (Portulaca oleracea). dandelion greens (Taraxacum spp.), watercress (Nasturtium officinale) and others.

are gaining popularity as functional foods, as they contain health beneficial bioactive compounds, vitamins, minerals or essential oils.

Other supplements with proven health benefits are the prebiotics and probiotics. The prebiotics are indigestible fibers that stimulate the intestinal microbiota in humans and animals. And the probiotics are viable microorganisms, mostly bacteria and some yeast, which are helping digestion and food transformation, with various beneficial traits for their host organisms. Although most studied probiotics are those for human and animal health, there is a special category of probiotics associated to plants, which bring beneficial attributes to these hosts. These plant probiotics are beneficial microorganism living in symbiotic relationship with the plant hosts or in free-living association with plants. They can act in different ways to increase plant development, to protect plant against different stress factors or increase their production of biologically active compounds

(de Souza Vandenberghe et al., 2017). Therefore, selected plant probiotics could be annlied as bioprotectants, biocontrollers, biofertilizers or biostimulants, especially in organic gardening. These plant probiotic microorganisms could be found on plant surfaces, as epiphytes, root surface, as rhizospheric microorganisms, or inside plant tissues, as endophytes (Whipps et al., 2008; Dutta & Bora, 2019). The contact between plants and microorganisms is made at the rhizosphere, endosphere and phillosphere level. Plant roots penetrate various layers of soil substrate in search of nutrients and water. During soil exploration, they encounter millions of different microorganisms and have developed advanced genetic and metabolic mechanisms to both recruit and defend against microorganisms. Root colonization involves a complex molecular communication between microorganism and roots. Attracted by root exudates, the microorganisms migrate towards roots via chemotaxis and may colonize the root surface, and soil aggregates that form around roots, or both. Best studied microbial colonizers of the root system, with beneficial effects on their hosts, are PGPR (plant growth promoting rhizobacteria) and mycorrhiza. Most beneficial interactions between plants and their symbionts begin at the rhizosphere level and should be considered the first application management of plant probiotics (Walker et al., 2020). The phillosphere is considered a hostile environment for microorganisms due to the limited nutrient sources, and UV radiation, but microorganisms found at this level were able to regulate various processes in plants, such as production of biologic active compounds

including organic acids, ascorbic acid (vitamin and diverse phenolic compounds. flavonoids and essential oils (Lindow & Leveau, 2002: Flores-Felix et al., 2008). Endophytes are more involved in plant metabolic activity than other plant associated microorganisms. Thanks to their intra-tissue colonization they have an intimate relationship with their host. Although plants are not any symptomatology revealing endophytic colonization, many processes in plants are improved (Fadiji & Babalola, 2020). This review focuses on various plant probiotics and other plant associated microorganisms with beneficial traits for their host. Mostly, the role of plant associated microorganisms in the bioactive compounds synthesis in their hosts.

MATERIALS AND METHODS

This review, is presenting plants probiotics effect, as single population or consortia of microorganisms, on bioactive compounds production in their hosts.

Data collected in this study derives from research articles published since 1995, featuring the importance of plant associated microorganisms for their hosts (Table 1).

The beneficial aspects documented in this review are focused on the improved plant quality, regarding their nutritional and medicinal value, if properly colonized by certain microorganisms. The most studied bioactive compounds are the antioxidants, especially ascorbic acid (vitamin C); flavonoids, such as anthocyanin; chlorophyll; alkamides and essential oils.

Table 1. Plant probiotics used in agriculture
(adapted after Jiménez-Gómez et al., 2017)

Inoculated plants	Microbial inoculants & Plant	Triggered effects on	Reference
	probiotics	bioactive compound	
		composition	
Begonia malabarica	Glomus mosseae,	Increased various secondary	Selvaraj et al., 2017
	Bacillus coagulans,	metabolites, such as total	
	Trichoderma viridae	phenols, ortho-dihydroxy	
		phenols, tannins, flavonoids,	
		and alkaloids	
Borago officinalis	AMF inoculated soil	Increased content of	Rahimi et al., 2017
		carotenoids and chlorophyll	

Inoculated plants	Microbial inoculants & Plant probiotics	Triggered effects on bioactive compound	Reference
	probletics	composition	
Brassica oleracea	Bacillus megaterium, Pantoea agglomerans and B. subtilis	Increase chlorophyll content	Turan et al., 2014
Capsicum annuum	Rhizobium leguminosarum PETP01	Increased antioxidant activity	Silva et al., 2014
Cymbopogon citratus	Azotobacter sp. and Pseudomonas sp.	Increased total flavonoid and total phenol content, improved the antioxidant capacity of the essential oils and helped lemongrass plants to tolerate better the abiotic stress	Mirzaei et al., 2020
Echinacea purpurea	Glomus intraradices	Increased production of secondary phytomedicinal metabolites	Araim et al., 2009
	Bacterial endophytes of <i>E. purpurea</i>	Increased concentration of alkamides	Maggini et al., 2017
Fagopyrum esculentum	Azospirillum spp. and Azotobacter spp. inoculants applied to buckwheat plants	Increased grain yield and concentrations of total flavonoid and phenol content in buckwheat	Singh et al., 2015
Fragaria x ananassa	Bacillus sp. RC23, B. cereus RC18, B. megaterium RC01 or Paenibacillus polymyxa RC05	Vitamin C enhancement	Erturk et al., 2012
	Phyllobacterium sp. PEPV15		Flores-Félix et al., 2015
Hibiscus sabdariffa	AMF or PGRP	Increased content of total chlorophyll and carotenoids in roselle	Sanayei et al., 2021
Hyoscyamus niger	Pseudomonas putida 168 or P. fluorescens 187 strains	Simulated antioxidant enzymes activity, increased proline accumulation, improved tropane alkaloid production and yield of root and shoot organs.	Ghorbanpour et al., 2013
Lactuca sativa	Endophytic plant growth promoting selenobacteria with or without mycorrhizal fungi	Increased content of total chlorophyll and carotenoids in lettuce	Durán et al., 2016
Lycopersicon esculentum	Bacillus amyloliquefaciens FZB42	Vitamin C enhancement	Gül et al., 2008
esemenum	Bacillus amyloliquefaciens and B. megaterium		Shen et al., 2016
	Pseudomonas sp. 19Fv1T		Bona et al., 2017
	Mixture of PGPR (Pseudomonas putida 41, Azotobacter chroococcum 5, and Azospirillum lipoferum OF strains) and AMF (Glomus intaradics, G. mossea, and G. etanicatum)	Increased antioxidant activity and lycopene content	Ordookhani et al., 2010
	Mixture of PGPR (A. chroococcum 5, and A. lipoferum OF strains) and AMF (G. intaradics, G. mossea, and G. etanicatum) Mixture of the PGPR strains: P. putida 41, A. chroococcum 5, and A. lipoferum OF strain		
	Bacillus licheniformis	Improved total flavonoids content	Ochoa-Velasco et al., 2016
Mentha piperita	Pseudomonas fluorescens	Increased essential oils amount	Banchio et al., 2008

Inoculated plants	Microbial inoculants & Plant probiotics	Triggered effects on bioactive compound composition	Reference
Mentha piperita	Plant growth promoting rhizobacteria	Increased amount of essential oils	Santoro et al., 2011
Ocimum basilicum	Bacillus subtilis GB03	Elevated α-terpenol and eugenol accumulation	Banchio et al., 2009
	Mixture of P. putida 41 strain, A. chroococcum 5 strain, A.lipoferum OF strain	Increased levels of essential oils antioxidant activity and microelements content	Ordookhani, 2011
	Bacillus lentus, Pseudomonas sp., Azospirillum brasilense	Increased antioxidant activity and chlorophyll leaf content	Heidari & Golpayegani, 2012
	Diazotrophs	Increased carotenoids and chlorophyll content in purple basil	Mariotti et al., 2021
Origanum majorana	Pseudomonas fluorescens Bradyrhizobium sp.	Increased amount of essentials oils	Banchio et al., 2008
Nasturtium officinale	Bacillus subtilis	Increased antioxidant capacity and total phenols of watercress	Pignata et al., 2016
Pelargonium graveolens	Bacillus subtilis, Pseudomonas fluorescens	Enhanced essential oil yield	Mishra et al., 2010
Pelargonium species	Glomus intraradices AMF and phosphate solubilizing bacteria	Enhanced yield and composition of essential oil (citronellol, geraniol, geranial, and eudesmol) in rose-scented geranium	Prasad et al., 2012
Ribes nigrum	ProbioHumus (Baltic Probiotics, Latvia) based on Saccharomyces cerevisiae yeast, Bacillus subtilis sporulated bacteria, Bifidobacterium animalis, B. bifidum, B. longum, Lactobacillus casei, L. diacetylactis, L. delbrueckii, L. plantarum, Lactococcus lactis, Streptococcus thermophiles lactic acid bacteria, Rhodopseudomonas palustris, and R. sphaeroides phototropic bacteria NaturGel containing also microorganisms of Azotobacter, Bacillus, Rhizobium, Bradyrhizobium, Lactobacillus, and Trichoderma genera Probiotic strains and	Ascorbic acid and anthocyanins increased content Enhanced antioxidant activity	Jurkonienė et al., 2021 Lingua et al., 2013
	mycorrhizal fungi	of berries and increased level of anthocyanins in fruits	Lingua et al., 2013
Rubus sp.	P. fluorescens N21.4	Increase and stabilize total flavonoid content in blackberry	Ramos-Solano et al., 2014
Spinacia oleracea	Rhizobium sp. PEPV12	Increase chlorophyll content	Jiménez-Gómez et al., 2016
Stevia rebaudiana	PGPR (such as Azotobacter chroococcum, Bacillus polymixa, Pseudomonas putida) with or without Glomus intraradices AMF	Increased chlorophyll content and augmented stevioside amount	Vafadar et al., 2014
Tagetes minuta	Pseudomonas fluorescens, Azospirillum brasilense	Increased monoterpene and phenolic compounds	del Rosario Cappellari at al., 2013

RESULTS AND DISCUSSIONS

The interaction between medicinal plants and their endophytic community was studied in order to review the increased production of biologic active compounds in microbial fortified plants. A significant increase of vitamin C level in plants was observed if microbial inoculated. The vitamin C is also known as ascorbic acid.

The results obtained by Flores-Félix et al. (2015) on strawberries showed that vitamin C levels were significantly higher in the fruits of plants inoculated with *Phyllobacterium* sp. PEPV15 strain, with approximately 79% more, compare to the uninoculated control. Previous studies performed by Erturk et al. (2012) showed high levels of vitamin C content in strawberry fruits after plants were inoculated with Paenibacillus polymyxa RC05 strain, as well as with Bacillus megaterum RC01, Bacillus cereus RC18 and Bacillus RC23 single strains, but not significant when treated with Bacillus RC03. Less significant differences regarding vitamin C content were observed in strawberry after foliar, and/or root application of Pseudomonas BA-8, Bacillus OSU-142 and Bacillus M-3 strains (Pirlak et al., 2009). Although these studies are revealing a wide variation regarding vitamin C levels in fruits of microbial inoculated strawberry plants, all bacteria used stimulated plant growth and increased different parameters of growth and productivity. This means that proper inoculants should be properly selected in order to increase the levels of bioactive compounds in plants.

Beside strawberries, blackcurrant berries are also known for their beneficial impact on human health (Hannum, 2004; Gopalan et al., 2012). Blackcurrant berries are rich anthocyanins. polyphenolic substances. antioxidants, vitamin C and gamma-linolenic acid. A recent study performed on blackcurrant grown in organic farming system showed that commercial plant probiotic products, like ProbioHumus and NaturGel increased the contents of ascorbic acid and anthocyanins. Moreover, when sprayed as single treatments or in combination, they significantly improved fruit yields with 38, 25, and 16%, respectively, compared with uninoculated (Jurkonienė et al., 2021).

Increased levels of vitamin C are also detected in vegetables after bacterial inoculation. In tomato fruits, the vitamin C content varies depending on production system (autumn or spring), nutrient system (open or closed-loop fertigation), concentrations of nutrient solution (full or half amount) and microbial treatments (PGPR inoculated or control). For instance, Gül et al. (2008) obtained best levels of vitamin C in tomato fruits (19.43 mg/100 ml) produced in spring growing season, when plants grown in perlite pots were inoculated with Bacillus in closed-loop amyloliquefaciens FZB42. fertigation system, with half amount of the normally used concentration of nutrient solution. Additionally, it was shows that vermicompost combined with plant probiotics Bacillus megaterium and B. amyloliquefaciens inoculation also increases vitamin C contents in fruits and tomato yield (Shen et al., 2016). Inoculation with *Pseudomonas* sp. 19Fv1T strain not only enhanced yield of tomato plants but also increased vitamin C concentration in fruits compared with the control treatment (Bona et al., 2017).

Beside vitamin C content, the antioxidant activity was also analyzed. On basil (Ocimum basilicum L.) studies were performed with various mixtures of Pseudomonas putida 41, Azotobacter chroococcum 5, and Azospirillum lipoferum OF strains presented increased levels of essential oils antioxidant activity and microelements content, compared to the control treatment (Ordookhani, 2011). Inoculants based on Pseudomonas sp., Bacillus lentus and Azospirillum brasilense on water stressed basil also increased antioxidant activity. Catalase and guaiacol peroxidase activity were higher as well as leaf chlorophyll content (Heidari & Golpayegani, 2012). The antioxidant activity was higher also in tomato fruits when plants were treated with the PGPR mixture of Pseudomonas putida 41. Azotobacter chroococcum 5, and Azospirillum lipoferum OF strains, with and without AMF (arbuscular mycorrhizal fungi) colonized soil (Ordookhani et al., 2010).

Bacterial inoculation with *Bacillus subtilis* increased the antioxidant capacity and total phenols of watercress (*Nasturtium officinale* R. Br.) medicinal plant (Pignata et al., 2016)

The antioxidant activity of berries can also be increased by plant microbial inoculation. Probiotic strains and mycorrhizal fungi are able to increase the level of anthocyanins in fruits (Lingua et al., 2013).

In addition to vitamin production and antioxidant activity, some plant probiotics are able to stimulate also carotenoids production. Such pigments are involved in photosynthesis. photoprotection, and act as stress hormones and signaling molecules in plants (Shumskaya & Wurtzel, 2013). Carotenoids are also revealing human health. nutrition and wellbeing attributes and some are precursors of vitamin A or act as antioxidants (Park et al., 2017). Beside the very well-known beta-carotene, lycopene is another known carotenoid. In tomato fruits, lycopene amount can be increased by proper PGPR inoculation of tomato plants. The combined treatment of PGPR and AMF on tomatoes also revealed higher levels of lycopene and potassium in tomato fruits (Ordookhani et al., 2010). Plants inoculation with Pseudomonas putida 41, Azotobacter chroococcum 5, and Azospirillum lipoferum OF strains of PGPR along with a mixture of Glomus lipoferum, G. mossea and G. etunicatum AMF increased not only lycopene amount in fruits but also antioxidant levels and potassium content in fruits and shoots (Ordookhani et al., 2010).

Carotenoids and chlorophyll content was increased in lettuce plants inoculated with endophytic growth promoting plant selenobacteria with or without mycorrhizal fungi (Durán et al., 2016). Similar effects were seen also in the medicinal plants Borago officinalis grown in AMF inoculated soil (Rahimi et al., 2017), in roselle (Hibiscus sabdariffa) inoculated either with AMF or PGRP (Sanayei et al., 2021), in purple basil (Ocimum basilicum L. cv. Red Rubin) inoculated with diazotrophs (Mariotti et al., 2021).

In Stevia rebaudiana, not only the chlorophyll content was increased but also the stevioside amount was augmented in biofertilised plants. Best results were obtained with the combined treatment of Azotobacter chroococcum PGPR + Glomus intraradices AMF, followed by other mixed treatments, such as Bacillus polymyxa +

G.intraradices, or A. chroococcum + Pseudomonas putida. Although, the other nine microbial biofertilizers also revealed improved NPK, chlorophyll and steviosede content compared to the untreated control (Vafadar et al., 2014).

Azospirillum spp. and Azotobacter spp. inoculants applied to buckwheat plants (Fagopyrum esculentum) increased grain yield and concentrations of total flavonoid and phenol content (Singh et al., 2015). In water stressed lemongrass (Cymbopogon citratus), Azotobacter sp. and Pseudomonas sp. inoculants not only that increased total flavonoid and total phenol content, but also improved the antioxidant capacity of the essential oils and helped plants to tolerate better the abiotic stress (Mirzaei et al., 2020).

Pseudomonas fluorescens N21.4 inoculation in *Rubus* sp. var. Lochness increase and stabilize total flavonoid content in blackberry (Ramos-Solano et al., 2014).

Effect of root colonization with selected microorganisms was studied to different medicinal plants in order to determine the composition and amount of essential oils. Pseudomonas fluorescens inoculation peppermint (Mentha piperita), as well as P. fluorescens + Bradyrhizobium sp. inoculation of oregano (Origanum majorana) increased total essential oil content without modifying the composition (Banchio et al., 2008). Later effect studies on microorganisms' peppermint showed that the volatile organic compounds of rhizobacteria can increase biosynthesis of essential oils and plant growth parameters (Santoro et al., 2011).

Phytosterols are essential biologic active compounds present in high concentrations in vegetable oils (Granado et al., 1995). Some studies suggest that application of plant probiotics can increase the levels of sterols in plants. Silva et al. (2014) inoculated two strains of *Rhizobium* (TVP08 and PEPT01) in pepper (*Capsicum annuum*) and evaluated their effect on sterols. The rhizobia inoculation produced a positive effect on the ripening of the pepper fruit, in addition to an improvement in several primary and secondary metabolites, which improved the nutritional value of the plant. Moreover, the aqueous extracts of *Capsicum*

annuum leaves, after bacterial inoculation with Rhizobium TVP08 strain, presented a significant acetylcholinesterase (AchE) inhibitory activity, with an important relevance in the treatment of Alzheimer's disease (Silva et al., 2014).

Alkamides from *Echinacea* have significant anti-inflammatory and immunomodulatory properties. Maggini et al. (2017) reported an increased concentration of alkamides in bacterial colonized plants of *Echinacea purpurea*.

Microbial inoculants perspectives

Studies on microbial inoculants are continuous process, either for new strains selection, analyzing their mechanisms of action, or understanding their implications on plants or environment. Microbial inoculants development is taking place at a global level. In agriculture, most of them are used as microbial fertilizers, biostimulants or plant protection However, in most countries. commercialization depends specific established standards. and government approval. However, according to policy initiatives addressed at the global level (e.g. Horizon 2020), it is absolutely necessary that the quantities of chemical fertilizers used be kept to a minimum and that "green products" be introduced (García-Fraile et al., 2015). Many countries have developed government policies for the use of biofertilizers, and biostimulants, although, the regulations are still in progress (García-Fraile et al., 2017).

In this context, collaboration between researchers and industry is a key factor in establishing the basis of a global biofertilizer market (García-Fraile et al., 2015).

Microbial inoculants or plant probiotics must meet a number of characteristics, but most importantly they must be safe for the environment and humans. Another important feature is their stability, and viability, also in terms of survival in certain biotic or abiotic conditions. Inoculants should include the most effective microbial strains that promote plant growth and productivity (Glick, 2012). Selected microbial strains should survive in wide range environmental conditions, as well as in certain abiotic stress factors (Chauhan et al., 2015). Various formulation types are available for

microbial inoculant used in agriculture most commons being the granules, powders, liquids, suspensions, emulsions and effervescent products (Kamilova et al., 2015; Lesueur et al., 2016; Zamfiropol-Cristea et al., 2017; Macik et al., 2020).

CONCLUSIONS

Microbial inoculants are having an important role in agriculture, stimulating plant growth and development. Their beneficial traits were demonstrated in both normal and stressful environmental conditions of plant production. This review documented important knowledge regarding microbial inoculants and their role in plant stimulation to produce phytomedicinal metabolites and nutritional compounds.

Such microbial inoculants can be considered plant probiotics, as they are able to improve plant physiological status in various environmental conditions. The applications of plant probiotics in agricultural practices enhance plants yield and quality.

REFERENCES

Araim, G., Saleem, A., Arnason, T., & Charest, C. (2009). Root colonization by arbuscular mycorrhizal (AM) fungus increases growth and secondary metabolism of purple cone flower, *Echinacea purpurea* (L.) Moench. *Journal of Agricultural and Food Chemistry*, 57 (6), 2255-2258. DOI: 10.1021/jf803173x.

Banchio, E., Bogino, P.C., Zygadlo, J., & Giordano, W. (2008). Plant growth promoting rhizobacteria improve growth and essential oil yield in *Origanum majorana* L. *Biochemical Systematics and Ecology*, 36 (10), 766-771.

Banchio, E., Xie, X., Zhang, H., & Paré, P. W. (2009). Soil bacteria elevate essential oil accumulation and emissions in sweet basil. J. Agric. Food Chem. 57, 653-657. DOI: 10.1021/jf8020305.

Bona, E., Cantamessa, S., Massa, N., Manassero, P., Marsano, F., Copetta, A., Lingua, G., D'Agostino, G., Gamalero, E., & Berta, G. (2017). Arbuscular mycorrhizal fungi and plant growth-promoting pseudomonads improve yield, quality and nutritional value of tomato: a field study. *Mycorrhiza*, 27(1), 1-11. https://doi.org/10.1007/s00572-016-0727-y.

Chauhan, H., Bagyaraj, D.J., Selvakumar, G., & Sundaram, S. (2015). Novel plant growth promoting rhizobacteria - Prospects and potential. *Applied Soil Ecology*, 95, 38-53.

del Rosario Cappellari, L., Valeria Santoro, M., Nievas, F., Giordano, W., & Banchio, E. (2013). Increase of secondary metabolite content in marigold by inoculation with plant growth-promoting

- rhizobacteria. *Applied Soil Ecology*, 70, 16-22. https://doi.org/10.1016/j.apsoil.2013.04.001.
- de Souza Vandenberghe, L. P., Garcia, L., Rodrigues, C., Camara, M. C., de Melo Pereira, G. V., de Oliveira, J., & Soccol, C. R. (2017). Potential applications of plant probiotic microorganisms in agriculture and forestry. AIMS microbiology, 3(3), 629-648. https://doi.org/10.3934/microbiol.2017.3.629.
- Durán, P., Acuña, J.J., Armada, E., López-Castillo, O.M., Cornejo, P., Mora, M.L., & Azcón, R., (2016). Inoculation with selenobacteria and arbuscular mycorrhizal fungi to enhance selenium content in lettuce plants and improve tolerance against drought stress. Journal of soil science and plant nutrition, 16 (1), 211-225. http://dx.doi.org/10.4067/S0718-95162016005000017.
- Dutta, J., & Bora, U. (2019). Chapter 18 Rhizosphere microbiome and plant probiotics. In J.S. Singh (Ed.), New and Future Developments in Microbial Biotechnology and Bioengineering, 273-281. Elsevier, https://doi.org/10.1016/B978-0-12-818258-1.00018-2.
- Erturk, Y., Ercisli, S., & Cakmakci, R. (2012). Yield and growth response of strawberry to plant growthpromoting rhizobacteria inoculation. *Journal of Plant Nutrition*, 35(6), 817-826.
- Fadiji, A.E., & Babalola, O.O. (2020). Exploring the potentialities of beneficial endophytes for improved plant growth. Saudi Journal of Biological Sciences, 27 (12), 3622-3633.
- Flores-Félix, J. D., Velázquez, E., García-Fraile, P., González-Andrés, F., Silva, L. R., & Rivas, R. (2018). *Rhizobium* and *Phyllobacterium* bacterial inoculants increase bioactive compounds and quality of strawberries cultivated in field conditions. *Food research international (Ottawa, Ont.), 111*, 416-422. https://doi.org/10.1016/j.foodres.2018.05.059.
- Flores-Félix, J. D., Silva, L. R., Rivera, L. P., Marcos-García, M., García-Fraile, P., Martínez-Molina, E., Mateos, P. F., Velázquez, E., Andrade, P., & Rivas, R. (2015). Plants probiotics as a tool to produce highly functional fruits: the case of *Phyllobacterium* and vitamin C in strawberries. *PloS One, 10*(4), e0122281.
- García-Fraile, P., Menéndez, E., & Rivas, R. (2015).Role of bacterial biofertilizers in agriculture and forestry. AIMS Bioengineering, 2 (3),183-205.
- García-Fraile, P., Menéndez, E., Celador-Lera, L., Díez-Méndez, A., Jiménez-Gómez, A., Marcos-García, M., Cruz-González, X.A., Martínez-Hidalgo, P., Mateos, P.F., & Rivas, R. (2017). Chapter 6 Bacterial Probiotics: A Truly Green Revolution. In: Kumar V., Kumar M., Sharma S., Prasad R. (eds). *Probiotics and Plant Health* (pp. 131-162). Springer, Singapore. https://doi.org/10.1007/978-981-10-3473-2_6.
- Ghorbanpour, M., Hatam, M., & Khavazi, K. (2013). Role of plant growth promoting rhizobacteria on antioxidant enzyme activities and tropane alkaloid production of Hyoscyamus niger under water deficit stress. Turkish Journal of Biology, 37, 350-360. DOI:10.3906/biy-1209-12.

- Glick, B.R. (2012). Plant growth-promoting bacteria: mechanisms and applications. *Scientifica*, 963401. http://dx.doi.org/10.6064/2012/963401.
- Gopalan, A., Reuben, S.C., Ahmed, S., Darvesh, A.S., Hohmannb, J., & Bishayee, A. (2012). The health benefits of blackcurrants. *Food & Function*, 3, 795-809.
- Granado, J., Felix, G., & Boller, T. (1995). Perception of fungal sterols in plants (subnanomolar concentrations of ergosterol elicit extracelular alkalinization in tomato cells). *Plant Physiology*, 107(2), 485-490. https://doi.org/10.1104/pp.107.2.485.
- Gül, A., Kidoglu, F., Tüzel, Y., & Tüzel, I.H. (2008). Effects of nutrition and *Bacillus amyloliquefaciens* on tomato (*Solanum lycopersicum* L.) growing in perlite. *Spanish Journal of Agricultural Research*, 6(3), 422-429. https://doi.org/10.5424/sjar/2008063-335.
- Hannum S. M. (2004). Potential impact of strawberries on human health: a review of the science. Critical Reviews in Food Science and Nutrition, 44(1), 1-17. https://doi.org/10.1080/10408690490263756.
- Heidari, M., & Golpayegani, A. (2012). Effects of water stress and inoculation with plant growth promoting rhizobacteria (PGPR) on antioxidant status and photosynthetic pigments in basil (Ocimum basilicum L.). Journal of the Saudi Society of Agricultural Sciences, 11(1), 57-61.
- Jiménez-Gómez, A., Celador-Lera, L., Fradejas-Bayón, M., & Rivas, R. (2017). Plant probiotic bacteria enhance the quality of fruit and horticultural crops. AIMS microbiology, 3(3). 483-501. https://doi.org/10.3934/microbiol.2017.3.483.
- Jiménez-Gómez, A., Menéndez, E., Flores-Félix, J.D.,
 García-Fraile, P. Mateos, P.F., & Rivas, R. (2016).
 Effective Colonization of Spinach Root Surface by
 Rhizobium. In: F.González-Andrés & E. James (Eds.)
 Biological nitrogen fixation and beneficial plant-microbe interaction, 109-122, Springer International
 Publishing. https://link.springer.com/ chapter/
 10.1007/978-3-319-32528-6
- Jurkonienė, S., Mockevičiūtė, R., Jankauskienė, J., Jankovska-Bortkevič, E., Armalytė, G., & Gavelienė, V. (2021). Application of Commercial Plant Probiotics Improves the Berry Yield and Quality of Field-Grown Blackcurrant. ACS Agricultural Science & Technology, 1(6), 615-622. https://doi.org/10.1021/acsagscitech.1c00115.
- Kamilova, F., Okon, Y., de Weert, S., & Hora, K. (2015) Commercialization of microbes: Manufacturing, inoculation, best practice for objective field testing, and registration. In: Lugtenberg B. (Ed.). *Principles* of *Plant-Microbe Interactions*, 319-328. Springer, Cham. https://doi.org/10.1007/978-3-319-08575-3 33.
- Lesueur, D., Deaker, R., Herrmann, L., Bräu, L., & Jansa, J. (2016) The production and potential of biofertilizers to improve crop yields. In: Arora N., Mehnaz S., Balestrini R. (Eds.) *Bioformulations: for sustainable agriculture*. 71-92. Springer, New Delhi. https://doi.org/10.1007/978-81-322-2779-3_4.
- Lindow, S.E., & Leveau, J.H. (2002). Phyllosphere microbiology. Current Opinion in Biotechnology 13. 238-243.

- Lingua, G., Bona, E., Manassero, P., Marsano, F., Todeschini, V., Cantamessa, S., Copetta, A., D'Agostino, G., Gamalero, E., & Berta, G. (2013). Arbuscular mycorrhizal fungi and plant growth-promoting pseudomonads increases anthocyanin concentration in strawberry fruits (*Fragaria x ananassa* var. Selva) in conditions of reduced fertilization. *International Journal of Molecular Sciences*, 14(8), 16207-16225. https://doi.org/10.3390/ijms140816207.
- Macik, M., Gryta, A., & Frac, M. (2020). Biofertilizer in agriculture: An overview on concepts, strategies and effects on soli microorganism. Advances in Agronomy, 162, 31-76.
- Maggini, V., De Leo, M., Mengoni, A., Gallo, E. R., Miceli, E., Reidel, R. V. B., Biffi, S., Pistelli, L., Fani, R., Firenzuoli, F., & Bogani, P. (2017). Plantendophytes interaction influences the secondary metabolism in *Echinacea purpurea* (L.) Moench: an in vitro model. Scientific Reports, 7(1), 16924. https://doi.org/10.1038/s41598-017-17110-w.
- Mariotti, L., Scartazza, A., Curadi, M., Picciarelli, P., & Toffanin, A. (2021). *Azospirillum baldaniorum* Sp245 induces physiological responses to alleviate the adverse effects of drought stress in purple basil. Plants, 10, 1141. DOI: 10.3390/plants10061141.
- Mirzaei, M., Ladan Moghadam, A., Hakimi, L., & Danaee. E. (2020). The Plant growth promoting rhizobacteria (PGPR) improve plant growth, antioxidant capacity, and essential oil properties of lemongrass (*Cymbopogon citratus*) under water stress. *Iranian Journal of Plant Physiology* 10(2), 3155-3166.
- Mishra, R. K., Prakash, O., Alam, M., & Dikshit, A. (2010). Influence of plant growth promoting rhizobacteria (PGPR) on the productivity of *Pelargonium graveolens* L. Herit. *Recent Research in Science and Technology* 2, 53-57.
- Ochoa-Velasco, C.E., Valadez-Blanco, R., Salas-Coronado, R., Sustaita-Rivera, F., Hernández-Carlos, B., García-Ortega, S., & Santos-Sánchez, N.F. (2016). Effect of nitrogen fertilization and *Bacillus licheniformis* biofertilizer addition on the antioxidants compounds and antioxidant activity of greenhouse cultivated tomato fruits (*Solanum lycopersicum* L. var. Sheva). *Scientia Horticulturae*, 201, 338-345.
- Ordookhani, K. (2011). Investigation of PGPR on antioxidant activity of essential oil and microelement contents of sweet basil. *Advances in Environmental Biology*, *5*, 1114-1120.
- Ordookhani, K., Khavazi, K., Moezzi, A., & Rejali, F. (2010). Influence of PGPR and AMF on antioxidant activity, lycopene and potassium contents in tomato. *African Journal of Agricultural Research*, 5(10), 1108-1116.
- Park, Y. J., Park, S. Y., Valan Arasu, M., Al-Dhabi, N. A., Ahn, H. G., Kim, J. K., & Park, S. U. (2017). Accumulation of carotenoids and metabolic profiling in different cultivars of tagetes flowers. *Molecules (Basel, Switzerland)*, 22(2), 313. https://doi.org/10.3390/molecules22020313.

- Pignata, G., Niñirola, D., Casale, M., Lo Turco, P.E., Egea-Gilabert, C., Fernández, J.A., & Nicola, S. (2016).Inherent quality and safety of watercress grown in a floating system using *Bacillus subtilis*. *The Horticulture Journal*, 85(2), 148-153. doi: 10.2503/horti.MI-091.
- Pırlak, L., & Köse, M. (2009). Effects of plant growth promoting rhizobacteria on yield and some fruit properties of strawberry. *Journal of Plant Nutrition*, 32(7), 1173-1184.
- Prasad, A., Kumar, S., Pandey, A., & Chand, S. (2012). Microbial and chemical sources of phosphorus supply modulate the yield and chemical composition of essential oil of rose-scented geranium (Pelargonium species) in sodic soils. Biology and Fertility of Soils, 48(1), 117-122.
- Ramos-Solano, B., Garcia-Villaraco, A., Gutierrez-Mañero, F. J., Lucas, J. A., Bonilla, A., & Garcia-Seco, D. (2014). Annual changes in bioactive contents and production in field-grown blackberry after inoculation with *Pseudomonas fluorescens*. *Plant Physiology and Biochemistry*, 74, 1-8. https://doi.org/10.1016/j.plaphy.2013.10.029.
- Rahimi, A., Jahanbin, Sh., Salehi, A., & Farajee, H. (2017). Changes in content of chlorophyll, carotenoids, phosphorus and relative water content of medicinal plant of borage (*Borago officinails* L.) under the influence of mycorrhizal fungi and water stress. *Journal of Biological Sciences*, 17, 28-34. DOI: 10.3923/jbs.2017.28.34
- Sanayei, S., Barmaki, M., Ebadi, A., & Torabi-Giglou, M. (2021). Amelioration of water deficiency stress in roselle (Hibiscus sabdariffa) by arbuscular mycorrhizal fungi and plant growth-promoting rhizobacteria. Notulae Botanicae Horti Agrobotanici Cluj-Napoca, 49(2), 11987. DOI: 10.15835/nbha49211987.
- Santoro, M.V., Zygadlo, J., Giordano, W., & Banchio, E. (2011). Volatile organic compounds from rhizobacteria increase biosynthesis of essential oils and growth parameters in peppermint (Mentha piperita). Plant Physiology and Biochemistry, 49, 1177-1182.
- Selvaraj, T., Rajeshkumar, S., Nisha, M.C., Wondimu, L. & Tesso. M. (2008). Effect of Glomus mosseae and plant growth promoting rhizomicroorganisms (PGPR's) on growth nutrients and content of secondary metabolites in Begonia malabarica Lam. International Journal of Science and Emerging Technologies, 2, 516-525.
- Shumskaya, M., & Wurtzel, E.T. (2013). The carotenoid biosynthetic pathway: Thinking in all dimensions. *Plant Science*, 208, 58-63.
- Shen, F., Zhu, T-B., Teng, M-J., Chen, Y., Liu, M-Q., Hu, F., & Li, H-X. (2016). [Effects of interaction between vermicompost and probiotics on soil nronerty, yield and quality of tomato.] Ying Yong Sheng Tai Xue Bao, 27(2), 484-490.
- Silva, L. R., Azevedo, J., Pereira, M. J., Carro, L., Velazquez, E., Peix, A., Valentão, P., & Andrade, P. B. (2014). Inoculation of the nonlegume *Capsicum annuum* (L.) with *Rhizobium* strains. 1. Effect on bioactive compounds, antioxidant activity, and fruit

- ripeness. Journal of Agricultural and Food Chemistry, 62 (3), 557-564. DOI: 10.1021/jf4046649.
- Singh, R., Babu, S., Avasthe, R.K., Yadav, G. S., Chettri, T.K., Phempunadi, C. D. & Chatterjee, T. (2015). Bacterial inoculation effect on soil biological properties, growth, grain yield, total phenolic and flavonoids contents of common buckwheat (Fagopyrum esculentum Moench) under hilly ecosystems of North-East India. African Journal of Microbiology Research, 9(15), 1110-1117.
- Turan, M., Ekinci, M., & Yildirim, E. (2014). Plant growth-promoting rhizobacteria improved growth, nutrient, and hormone content of cabbage (*Brassica oleracea*) seedlings. *Turkish Journal of Agriculture and Forestry*, 38, 327-333.
- Vafadar, F., Amooaghaie, R., & Otroshy, M. (2014). Effects of plant-growth-promoting rhizobacteria and

- arbuscular mycorrhizal fungus on plant growth, stevioside, NPK, and chlorophyll content of *Stevia rebaudiana*. *Journal of Plant Interactions*, *9* (1), 128-136. DOI: 10.1080/17429145.2013.779035.
- Walker, R., Otto-Pille, C., Gupta, S., Schillaci, M., & Roessner, U. (2020). Current perspectives and applications in plant probiotics. *Microbiology Australia* 41. 95-99. DOI: 10.1071/MA20024.
- Whipps, J.M., Hand, P., Pink, D., & Bending, G.D. (2008). Phyllosphere microbiology with special reference to diversity and plant genotype, *Journal of Applied Microbiology*, 105, 1744-1755.
- Zamfiropol-Cristea V., Răut I., Şeşan T.E., Trică B., & Oancea F. (2017). Surface response optimization of submerged biomass production for a plant biostimulant Trichoderma strain. *Scientific Bulletin. Series F. Biotechnologies*, 21, 56-65.